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1       **Direct or indirect regulation of muscle protein synthesis by energy status?**

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16      **Conflict of interest:** C. Moinard is a shareholder in Citrage. Eric Fontaine has no conflict of interest to  
17      report.

18

19

20 **Abstract:**

21 Muscle protein synthesis (MPS) is a complex and finely-regulated mechanism that plays a key role in  
22 muscle homeostasis. Amino acid bioavailability is widely considered a major driver of MPS regulation  
23 via mTOR pathway activation. However, recent results suggest that amino acid bioavailability affects  
24 cellular energy status. Whatever the tool used to modulate energy status (amino acid depletion or  
25 mild mitochondrial uncoupling), a decrease in cellular energy status decreases MPS, without  
26 necessarily involving the mTOR pathway. Here we propose that energy status directly regulates one  
27 or several energy-consuming step(s) during MPS. This new paradigm modifies our vision of protein  
28 metabolism and raises prospects for new advances in therapeutics.

29

30 **Introduction:**

31 Human skeletal muscle is the main store of proteins at whole-body level. Skeletal muscles  
32 drive the locomotor function but they also have an important metabolic role. Muscle protein  
33 turnover (i.e. protein synthesis and proteolysis) is a tightly regulated process. Muscle growth occurs  
34 when protein synthesis is higher than proteolysis, whereas muscle atrophy occurs when protein  
35 synthesis is lower than proteolysis [1].

36 New proteins get synthesized through multiple reactions that occur in the nucleus, the  
37 cytosol, or various subcellular locations [2]. The process begins by mRNA translation, which proceeds  
38 in 3 phases: initiation, elongation, and termination. The level of muscle protein synthesis is governed  
39 by two factors: translational efficiency (i.e. the rate of protein synthesized by ribosome) and  
40 translational capacity (i.e. the number of ribosomes/units of tissue; see [3] for an extensive review).  
41 Crucially, these different steps demand a large amount of energy to drive both chemical reactions  
42 (such as peptide bonds and phosphorylation) and mechanical reactions (ribosome displacements) [4].  
43 Protein synthesis is thought to be mainly, if not totally, under control of the mTOR signaling pathway,  
44 which is where both exogenous (i.e. amino acids) and endogenous (i.e. AMPK) stimuli converge  
45 (Figure 1). Based on the effects of the mTORC1 inhibitor rapamycin, there is no doubt that mTORC1  
46 regulates protein synthesis [3]. There is ample literature showing that mTORC1 can regulate several  
47 steps in protein synthesis, [1-3, 5, 6] but the molecular regulatory mechanisms involved are not  
48 always fully understood.

49

50 **Regulation of muscle protein synthesis by amino acids**

51 Exogenous regulatory factors include amino acids and hormones, which are recognized as  
52 major regulators of protein synthesis. After a meal, there is an increase in amino acid availability at  
53 peripheral level associated with an increase of insulin, both of which are required to stimulate MPS

54 *via* direct activation of the mTORC1 pathway [5, 6]. A study using a model of isolated perfused  
55 muscle showed that maximal stimulation occurred with the normal postprandial amino acid  
56 concentrations [7], and *in vitro* studies using myotube cultures also found that high levels of amino  
57 acids are associated with high levels of protein synthesis [8].

58         From a finalist point of view, it seems logical that amino acids stimulate protein synthesis,  
59 and especially muscle synthesis. Indeed, the organism has to fight on two fronts. On one hand, there  
60 is no storage capacity for amino acids (unlike glycogen which has no other function than the storage  
61 of glucose, proteins are primarily synthesized to perform a function). On the other hand, amino acids  
62 are neurotoxic and so the body needs to limit any excessive hyperaminoacidemia (amino acids are  
63 eliminated via the urea cycle).

64

#### 65 **Regulation of muscle protein synthesis by energy level**

66         In most cells, ATP is mainly produced by mitochondria during a process that couples oxygen  
67 consumption to ATP production (OXPHOS machinery). Because the fluxes of ATP production and ATP  
68 consumption are not directly measurable in a living cell, oxygen consumption is a surrogate of ATP  
69 consumption in intact cells, tissues, and organisms.

70         It is assumed that the inhibition of an ATP-consuming process results in a decrease in cellular  
71 oxygen consumption. In practice, this reduction in oxygen consumption is measurable when a major  
72 metabolic pathway is inhibited. It has been shown under controlled conditions that during the  
73 gradual inhibition of mitochondrial respiration, some ATP-consuming processes decrease rapidly (the  
74 inhibition of the metabolic pathway has virtually no effect on remaining oxygen consumption) while  
75 all the others are maintained [9]. These observations led to the conclusion that there is a hierarchy  
76 between the various processes that consume ATP in cells, with some stopping when ATP production  
77 decline and others persisting despite it [9]. Note that this experimental approach is based on two

88 assumptions: first, that all the ATP-consuming processes in a given pathway are inhibited; second,  
89 that other ATP-consuming processes are not increased. With these hypotheses fresh in mind, note  
90 that it has been reported that protein synthesis is strongly inhibited as soon as mitochondrial  
91 respiration is weakly decreased [9], while Na<sup>+</sup> cycling and Ca<sup>2+</sup> cycling are almost unaffected [9]. It is  
92 proposed that shutting down protein synthesis serves to save ATP for more immediately vital cells  
93 functions.

84 The concept of a hierarchy between several ATP-consuming processes in cells implies the  
85 existence of a signal capable of inhibiting certain metabolic pathways but not others. The progressive  
86 inhibition of mitochondrial respiration not only leads to gradual inhibition of ATP synthesis rate, it  
87 also results in a decrease in the ATP/ADP ratio.

88 By sensing the concentration of AMP, which is in equilibrium with the ATP/ADP ratio, the  
89 AMP-activated protein kinase (AMPK) is an important sensor of cellular energy status [10]. Given that  
90 AMPK can inactivate mTOR, it has been proposed that energy status controls protein synthesis via its  
91 effect on AMPK. Although this proposal cannot be ruled out, recent data are not consistent with a  
92 pure effect of energy status on mTOR, which leads us to propose that besides signaling regulation,  
93 protein synthesis is also directly thermodynamically regulated by the ATP/ADP Ratio.

94 Using a model of primary myotubes, we evaluated the effect of amino acid starvation on MPS  
95 (evaluated by the Sunset method), and energy status [4]. As expected, amino acid starvation led to a  
96 decrease in MPS (-29%), the remaining MPS being made from endogenous amino acids. Surprisingly,  
97 we also observed that amino acid starvation led to a decrease in energy status (ATP/ADP ratio) (-  
98 13%) and to an AMPK activation. In order to distinguish the respective role of these two regulators  
99 (amino acid starvation and energy status) in decreasing MPS, we used a model of myotubes with a  
100 reduced cellular energy status induced by mild uncoupling of the OXPHOS machinery. We observed  
101 that the same decrease in ATP/ADP ratio led to a decrease in protein synthesis (-21%) despite the

102 presence of a high concentration of amino acids in the medium. At this stage, these results remained  
103 in line with the proposition that MPS was decreased due to mTOR regulation. We then tested the  
104 effect of leucine or citrulline, which **both** activate MPS in cases of amino acid deficiency. We  
105 observed that citrulline but not leucine was able to counteract the inhibition of MPS. Importantly,  
106 citrulline did not normalize the ATP/ADP ratio and had no effect on AMPK. In other words, citrulline  
107 has no effect on mTOR but increases MPS under energy stress. Importantly, however, citrulline did  
108 not increase MPS under control conditions, indicating that citrulline as such is not a direct activator  
109 of MPS.

110

#### 111 **ATP for paying, not for signaling**

112 Citrulline only stimulates MPS when there is energy stress, which it does not improve. To  
113 account for the fact, we propose a regulation of protein synthesis based on thermodynamic  
114 considerations. Thermodynamics laws stipulate that a reaction can only take place when it is  
115 accompanied by a decrease in its Gibbs' free energy (i.e. when the  $\Delta G$  of the reaction is  $< 0$ ).  
116 However, non-thermodynamically favorable reactions ( $\Delta G > 0$ ) can occur if they are coupled with  
117 thermodynamically favorable reactions ( $\Delta G < 0$ ) as long as the sum of the two  $\Delta G$  remains  $< 0$ . As a  
118 rule, the reaction that “pays for” a non-thermodynamically favorable reaction is the hydrolysis of ATP  
119 into ADP plus Pi (phosphate).

120 Remember that the direction of a reversible reaction depends on the respective  
121 concentrations of the reagents and the products of the reaction. This phenomenon, known as the  
122 “law of mass action”, means that the  $\Delta G$  of the reaction can move from a negative value (when the  
123 reaction occurs in the forward direction) to a positive value (when the reaction occurs in the reverse  
124 direction). In other words, the  $\Delta G$  of a given reaction changes according to the concentrations of its  
125 reagents and products. In the case of ATP synthesis/hydrolysis ( $ATP \leftrightarrow ADP + Pi$ ), this means that the

126  $\Delta G$  of ATP hydrolysis depends on the concentrations of ATP, ADP and Pi. If we apply the law of mass  
127 action to this reaction, then the  $\Delta G$  of ATP hydrolysis depends on the  $\text{ATP}/(\text{ADP} \times \text{Pi})$  ratio. In other  
128 words, when a molecule of ATP is hydrolyzed, the energy available depends not only on the  
129 concentration of ATP but also on the concentrations of ADP and Pi. Physiologically, the concentration  
130 of Pi is much higher than the concentrations of ATP and ADP, so it is mainly the ATP/ADP ratio that  
131 influences the  $\Delta G$  of the reaction. By analogy with money, ATP molecules are like coins, whose value  
132 depends on the ATP/ADP ratio. This takes us to a counter-intuitive thermodynamic truth: it is not the  
133 availability of ATP that controls reaction, but its value (i.e. its ATP/ADP ratio).

134 Protein synthesis involves many energy-consuming steps hydrolyzing either ATP or GTP  
135 (which are at thermodynamic equilibrium *via* the nucleoside diphosphate kinase). It is thus likely that  
136 one or several step(s) require a large amount of energy that cannot be released by the hydrolysis of  
137 an ATP with a low ATP/ADP ratio (or a GTP with a low GTP/GDP ratio). In other words, the reaction  
138 does not take place because the energy required is insufficient.

139 Although no reaction (alone or coupled to an energy-releasing reaction) can be performed if  
140 its  $\Delta G$  is not negative, some reactions do not occur despite a negative  $\Delta G$ . More precisely, the  
141 reaction occurs but so slowly that we do not perceive it. Thermodynamically, this apparent paradox  
142 is explained by the existence of a "potential barrier" (sometimes called the "energy barrier").  
143 Importantly, the size of the potential barrier depends on the  $\Delta G$  (it increases when the  $\Delta G$  becomes  
144 less negative). To be overcome (i.e. so that the reaction occurs), this potential barrier requires an  
145 activation energy. In biology, the thermodynamically favorable reactions are regulated by enzymes  
146 that decrease the potential barrier of reactions. In other words, enzymes facilitate  
147 thermodynamically favorable reactions by decreasing their activation energy, but they are unable to  
148 catalyze a non-thermodynamically favorable reaction. We propose that citrulline can also reduce the  
149 potential barrier of one or more energy-consuming processes involved in protein synthesis.



150 Both thermodynamics and enzymes regulate metabolic pathways. However, the respective  
151 share of the thermodynamic component and the enzymatic (kinetic) component varies according to  
152 the reactions and, for a given reaction, with the conditions. In general, thermodynamic conditions  
153 control a reaction when its enzyme is activated or in high concentration, whereas enzymes become  
154 the main regulator when inhibited or in low concentration. This easily explains why citrulline does  
155 not stimulate protein synthesis under the control conditions (when ATP/ADP ratio is high) because,  
156 under these conditions, the potential barrier is already low.

157 It is abundantly clear that protein synthesis involves many enzymes or machineries that  
158 require energy consumption. All these steps are potentially controlled by signaling pathways (mTOR  
159 for example). However, it must be kept in mind that the activation of an enzyme has no effect if the  
160 energy required for the reaction is insufficient. We believe that protein synthesis can be regulated  
161 directly (thermodynamically) by the ATP/ADP ratio, independently of AMPK signaling. This  
162 thermodynamic regulation explains the observations of no activation of protein synthesis despite the  
163 activation of its signaling pathways.

164

#### 165 **In vivo data supporting our proposal.**

166 It is well known that malnourished patients lose muscle proteins and that this process  
167 contributes to morbidity and mortality (linked to time and intensity). Different strategies have been  
168 tested to limit protein loss, but with limited results. For example, using a rodent model of traumatic  
169 brain injury leading to muscle loss [11], we observed an impairment of energy status in tissues [12].  
170 We were able to normalize caloric and protein intake using enteral nutrition. **However**, this strategy  
171 did not restore energy status [12] nor **muscle protein content** [13]. **This observation is consistent**  
172 **with our proposal that the normalization of protein intake is not sufficient to restore muscle mass**  
173 **when energy status is impaired.**

174           Ageing induces a decrease of muscle mass by 50 % between 20 and 80 years old that can  
175 possibly lead to sarcopenia. Several pieces of evidence suggest that mitochondrial function is  
176 reduced during aging, both in muscle and in neurons (see [14, 15] for recent reviews). It is generally  
177 assumed that such dysfunction reduces physical activity, leading to muscle atrophy. Besides this  
178 mechanical hypothesis, a direct effect of energy status on protein synthesis has not yet been  
179 proposed but is supported by indirect evidence. Indeed, ageing leads to a decrease in ATP content  
180 [16, 17], while it decreases the rate of mitochondrial protein synthesis in muscle [18].

181           Several human studies concordantly conclude that a decrease in energy consumption leads  
182 to a decrease in the rate of protein synthesis [19-22]. Unfortunately, none of these studies measured  
183 energy status (the ATP/ADP ratio). Note however, that caloric restriction in rat led to a decrease in  
184 energy status (as measured by the activation of the AMPK pathway) [23]. Together, these results are  
185 consistent with our proposal that energy status directly regulates protein synthesis.

186

#### 187 **Relevance to clinical nutrition.**

188           Here, we propose that energy status is a major factor in the regulation of muscle protein  
189 synthesis. It acts indirectly via mTOR pathway signaling (regulation by AMPK) but also directly during  
190 the consumption of energy. From a theoretical point of view, two strategies could restore the  
191 synthesis of proteins when impaired by energy stress. The first is to restore energy homeostasis (i.e.  
192 to normalize the ATP/ADP ratio). The second is to use a protein synthesis catalyst capable of  
193 offsetting the effect of a low ATP/ADP ratio. For the time being, the first strategy is difficult to  
194 implement because the measurement of the energetic status is not done routinely. However, this is  
195 probably what happens during the healing of an acute disease. Alternatively, the use of citrulline (or  
196 other compounds that would act similarly) is a pragmatic approach **that could be implemented**. The  
197 development and systematic use of the measurement of the cellular energy status would be a

198 valuable tool to revisits our understanding of protein metabolism and lay the foundation for new  
199 approaches to the management of undernutrition.

200 **Conclusion**

201 In this article, we propose that the energy status (i.e., the ATP/ADP ratio) is bona fide  
202 regulator of protein synthesis. This proposal is supported by both direct and indirect evidence, but it  
203 remains to be confirmed in larger scale experiments in humans.

204

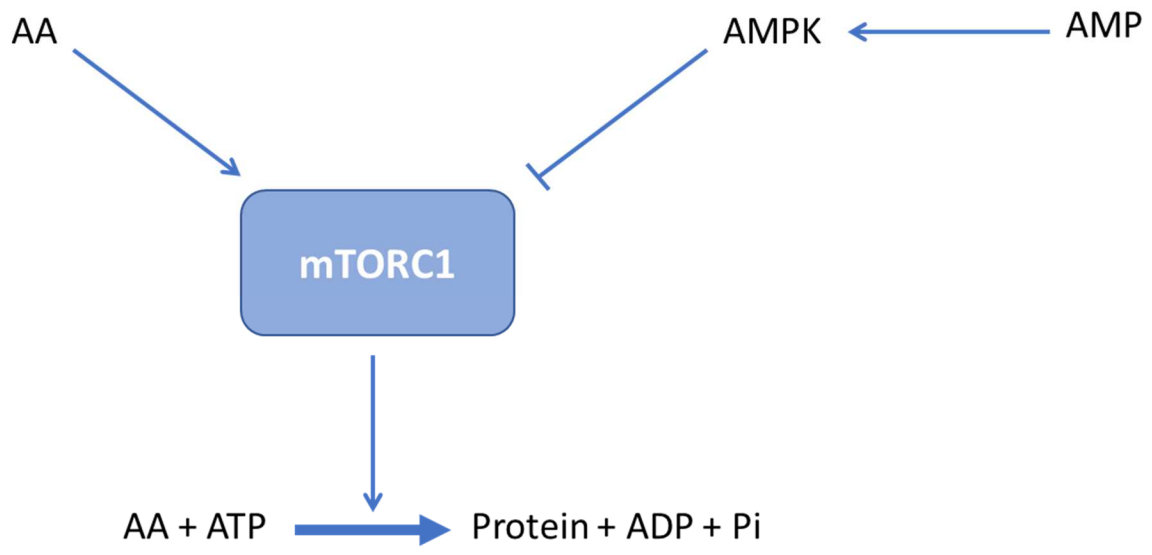
205

206 **References**

- 207 [1] Mirzoev TM, Shenkman BS. Regulation of Protein Synthesis in Inactivated Skeletal Muscle: Signal  
208 Inputs, Protein Kinase Cascades, and Ribosome Biogenesis. *Biochemistry (Mosc)*. 2018;83:1299-317.
- 209 [2] Prod'homme M, Rieu I, Balage M, Dardevet D, Grizard J. Insulin and amino acids both strongly  
210 participate to the regulation of protein metabolism. *Curr Opin Clin Nutr Metab Care*. 2004;7:71-7.
- 211 [3] Gordon BS, Kelleher AR, Kimball SR. Regulation of muscle protein synthesis and the effects of  
212 catabolic states. *Int J Biochem Cell Biol*. 2013;45:2147-57.
- 213 [4] Goron A, Lamarche F, Blanchet S, Delangle P, Schlattner U, Fontaine E, et al. Citrulline stimulates  
214 muscle protein synthesis, by reallocating ATP consumption to muscle protein synthesis. *J Cachexia*  
215 *Sarcopenia Muscle*. 2019;10:919-28.
- 216 [5] Balage M, Sinaud S, Prod'homme M, Dardevet D, Vary TC, Kimball SR, et al. Amino acids and  
217 insulin are both required to regulate assembly of the eIF4E. eIF4G complex in rat skeletal muscle. *Am*  
218 *J Physiol Endocrinol Metab*. 2001;281:E565-74.
- 219 [6] James HA, O'Neill BT, Nair KS. Insulin Regulation of Proteostasis and Clinical Implications. *Cell*  
220 *Metab*. 2017;26:310-23.
- 221 [7] Dardevet D, Sornet C, Balage M, Grizard J. Stimulation of in vitro rat muscle protein synthesis by  
222 leucine decreases with age. *J Nutr*. 2000;130:2630-5.
- 223 [8] Iresjo BM, Svanberg E, Lundholm K. Reevaluation of amino acid stimulation of protein synthesis in  
224 murine- and human-derived skeletal muscle cells assessed by independent techniques. *Am J Physiol*  
225 *Endocrinol Metab*. 2005;288:E1028-37.
- 226 [9] Buttgerit F, Brand MD. A hierarchy of ATP-consuming processes in mammalian cells. *Biochem J*.  
227 1995;312 ( Pt 1):163-7.
- 228 [10] Thomson DM. The Role of AMPK in the Regulation of Skeletal Muscle Size, Hypertrophy, and  
229 Regeneration. *Int J Mol Sci*. 2018;19.
- 230 [11] Moinard C, Neveux N, Royo N, Genthon C, Marchand-Verrecchia C, Plotkine M, et al.  
231 Characterization of the alteration of nutritional state in brain injury induced by fluid percussion in  
232 rats. *Intensive Care Med*. 2005;31:281-8.
- 233 [12] Moinard C, Gupta S, Besson V, Morio B, Marchand-Leroux C, Chaumeil JC, et al. Evidence for  
234 impairment of hepatic energy homeostasis in head-injured rat. *J Neurotrauma*. 2008;25:124-9.
- 235 [13] Charrueau C, Belabed L, Besson V, Chaumeil JC, Cynober L, Moinard C. Metabolic response and  
236 nutritional support in traumatic brain injury: evidence for resistance to renutrition. *J Neurotrauma*.  
237 2009;26:1911-20.
- 238 [14] Alway SE, Mohamed JS, Myers MJ. Mitochondria Initiate and Regulate Sarcopenia. *Exerc Sport*  
239 *Sci Rev*. 2017;45:58-69.
- 240 [15] Short KR, Nair KS. Does aging adversely affect muscle mitochondrial function? *Exerc Sport Sci*  
241 *Rev*. 2001;29:118-23.

- 242 [16] Conley KE, Jubrias SA, Esselman PC. Oxidative capacity and ageing in human muscle. *J Physiol.*  
243 2000;526 Pt 1:203-10.
- 244 [17] Drew B, Phaneuf S, Dirks A, Selman C, Gredilla R, Lezza A, et al. Effects of aging and caloric  
245 restriction on mitochondrial energy production in gastrocnemius muscle and heart. *Am J Physiol*  
246 *Regul Integr Comp Physiol.* 2003;284:R474-80.
- 247 [18] Rooyackers OE, Adey DB, Ades PA, Nair KS. Effect of age on in vivo rates of mitochondrial protein  
248 synthesis in human skeletal muscle. *Proc Natl Acad Sci U S A.* 1996;93:15364-9.
- 249 [19] Hector AJ, McGlory C, Damas F, Mazara N, Baker SK, Phillips SM. Pronounced energy restriction  
250 with elevated protein intake results in no change in proteolysis and reductions in skeletal muscle  
251 protein synthesis that are mitigated by resistance exercise. *FASEB J.* 2018;32:265-75.
- 252 [20] Mojtahedi MC, Thorpe MP, Karampinos DC, Johnson CL, Layman DK, Georgiadis JG, et al. The  
253 effects of a higher protein intake during energy restriction on changes in body composition and  
254 physical function in older women. *J Gerontol A Biol Sci Med Sci.* 2011;66:1218-25.
- 255 [21] Murphy CH, Churchward-Venne TA, Mitchell CJ, Kolar NM, Kassis A, Karagounis LG, et al.  
256 Hypoenergetic diet-induced reductions in myofibrillar protein synthesis are restored with resistance  
257 training and balanced daily protein ingestion in older men. *Am J Physiol Endocrinol Metab.*  
258 2015;308:E734-43.
- 259 [22] Pasiakos SM, Vislocky LM, Carbone JW, Altieri N, Konopelski K, Freake HC, et al. Acute energy  
260 deprivation affects skeletal muscle protein synthesis and associated intracellular signaling proteins in  
261 physically active adults. *J Nutr.* 2010;140:745-51.
- 262 [23] Jiang W, Zhu Z, Thompson HJ. Dietary energy restriction modulates the activity of AMP-activated  
263 protein kinase, Akt, and mammalian target of rapamycin in mammary carcinomas, mammary gland,  
264 and liver. *Cancer Res.* 2008;68:5492-9.
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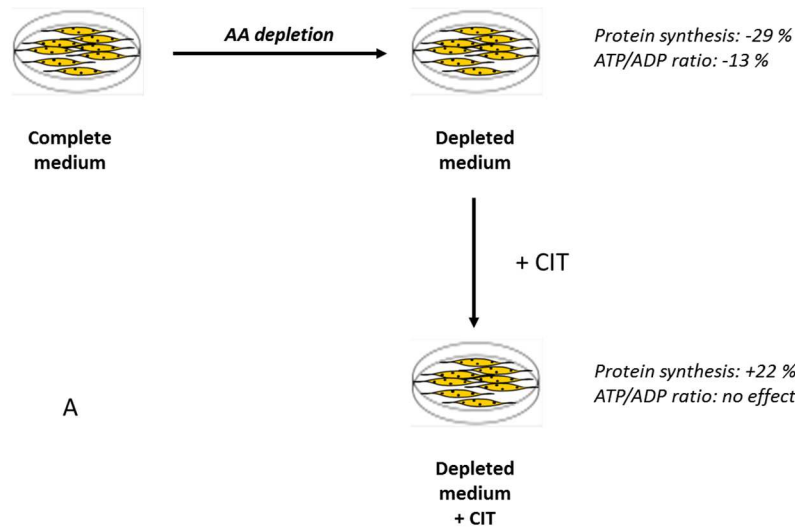
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269 **Figure 1:** General overview of regulation of protein synthesis by amino acids and energy status

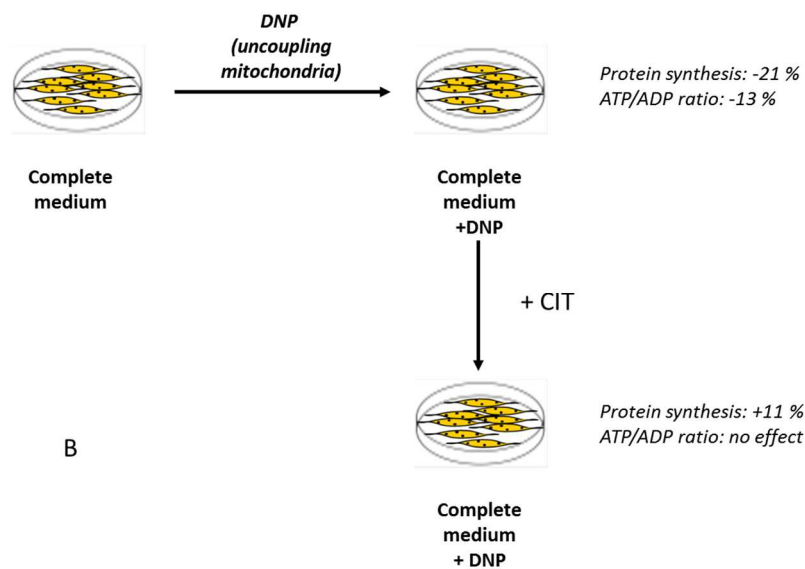
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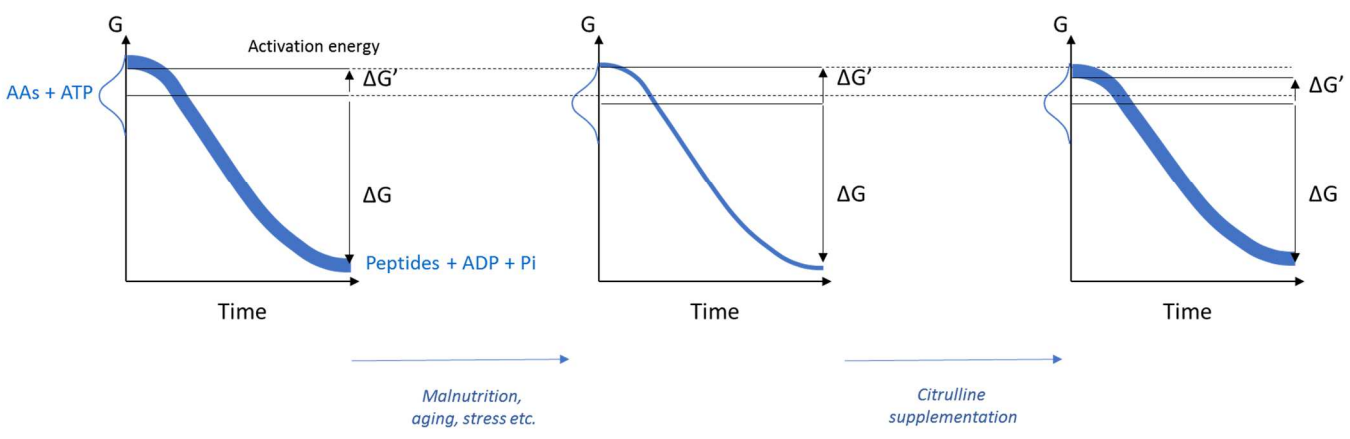
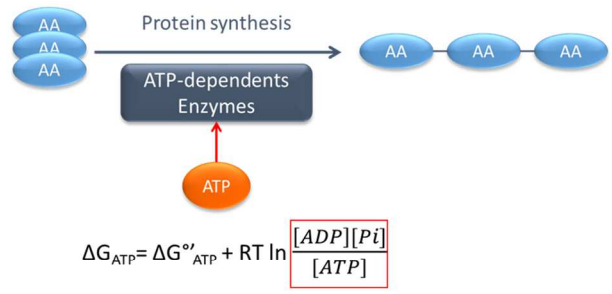
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275 **Figure 2:** In myotubes, AAs deficiency for 16h decreases both protein synthesis and energy status. In  
 276 these conditions, CIT is able to restore protein synthesis without modification of ATP/ADP ratio  
 277 (Figure 2A). In a second set of experiments (Figure 2B), cellular energy state was decreased by mildly  
 278 uncoupling mitochondria (as the same level as amino acid deficiency) but with a complete medium  
 279 (with complete AA medium). Such conditions also decrease protein synthesis and CIT again  
 280 stimulates protein synthesis (adapted from Goron et al., *J Cachexia Sarcopenia Muscle*, 2019).

281



282

283

284

285 **Figure 3:** At the molecular level, the G of reagents is a value that fluctuates around an average state.  
 286 Only reagents whose G exceeds the G of the activation energy are converted into product. As the  
 287 average G decreases, the number of reagents exceeding the G of the activation energy decreases. As  
 288 a result, the rate of reaction decreases. Thus, to accelerate a reaction, it is necessary to increase the  
 289 average G or to decrease the activation energy.

290 When applied to protein synthesis, ATP-consuming processes (or GTP-consuming processes) slow  
 291 down when the average G of ATP (or GTP) decreases. Citrulline would restore the initial velocity by  
 292 decreasing the activation energy.

293