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## Phospholipase D inhibitors screening: probing and evaluation of ancient and novel molecules

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<b>Abstract:</b>	Phospholipase D (PLD) is a ubiquitous enzyme that cleaves the distal phosphoester bond of phospholipids generating phosphatidic acid (PA). In plants, PA is involved in numerous cell responses triggered by stress. Similarly, in mammals, PA is also a second messenger involved in tumorigenesis. PLD is nowadays considered as a therapeutic target and blocking its activity with specific inhibitors constitutes a promising strategy to treat cancers. Starting from already described PLD inhibitors, this study aims to investigate the effect of their structural modifications on the enzyme's activity, as well as identifying new potent inhibitors of eukaryotic PLDs. Being able to purify the plant PLD from <i>Vigna unguiculata</i> (VuPLD), we obtained a SAXS model of its structure. We then used a fluorescence-based test suitable for high-throughput screening to review the effect of eukaryotic PLD inhibitors described in the literature. In this regard, we found that only few molecules were in fact able to inhibit VuPLD and we confirmed that vanadate is the most potent of all with an IC50 around 58 µM. Moreover, the small-scale screening of a chemical library of 3,120 compounds allowed us to optimize the different screening's steps and paved the way towards the discovery of new potent inhibitors.
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<b>Response to Reviewers:</b>	

# Phospholipase D inhibitors screening: probing and evaluation of ancient and novel molecules

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**Keywords:** phospholipase D, inhibitor, phosphatidic acid.

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21 **Abstract**

22 Phospholipase D (PLD) is a ubiquitous enzyme that cleaves the distal phosphoester bond of  
23 phospholipids generating phosphatidic acid (PA). In plants, PA is involved in numerous cell  
24 responses triggered by stress. Similarly, in mammals, PA is also a second messenger involved in  
25 tumorigenesis. PLD is nowadays considered as a therapeutic target and blocking its activity with  
26 specific inhibitors constitutes a promising strategy to treat cancers. Starting from already described  
27 PLD inhibitors, this study aims to investigate the effect of their structural modifications on the  
28 enzyme's activity, as well as identifying new potent inhibitors of eukaryotic PLDs. Being able to  
29 purify the plant PLD from *Vigna unguiculata* (VuPLD), we obtained a SAXS model of its  
30 structure. We then used a fluorescence-based test suitable for high-throughput screening to review  
31 the effect of eukaryotic PLD inhibitors described in the literature. In this regard, we found that  
32 only few molecules were in fact able to inhibit VuPLD and we confirmed that vanadate is the most  
33 potent of all with an IC<sub>50</sub> around 58 μM. Moreover, the small-scale screening of a chemical library  
34 of 3,120 compounds allowed us to optimize the different screening's steps and paved the way  
35 towards the discovery of new potent inhibitors.

## 36 Introduction

37 Phospholipase D (PLD) (EC 3.1.4.4) is a diverse family of enzymes found in microbes, fungi,  
38 viruses, plants and animals [1]. In addition to its hydrolytic activity towards phospholipids  
39 releasing phosphatidic acid (PA) and a polar moiety, PLD catalyzes a transphosphatidylation  
40 reaction as well. This reaction occurs when a primary alcohol is used instead of water, making  
41 possible the synthesis of rarer phospholipids by exchanging the polar headgroup, as  
42 phosphatidylserine from the abundant phosphatidylcholine (PC) in the presence of serine.

43 Contrary to several bacterial PLDs, plant and mammalian PLDs share a common feature  
44 represented by a duplicated HKD motif that is involved in the catalytic reaction, there are then two  
45 classes of PLDs based on their primary structure: the HKD-containing and the non-HKD [2]. This  
46 latter family regroups well-used enzymes that belongs to the superfamily of PLC-like  
47 phosphodiesterases [3], such as the commercial PLD from *Streptomyces chromofuscus* [4] or the  
48 PLD from arachnids [5]. The HKD-containing PLD family belongs to the PLD superfamily and is  
49 mostly found in mammals as for example PLD1 and PLD2 in *Homo sapiens*, in plants as PLD $\alpha$ ,  
50 in yeast with SPO14 [2,6,7], and in several microbes as the crystallized PLD from *Streptomyces*  
51 *sp.* strain PMF[8]. The hallmark of the PLD superfamily is the presence of the so-called HKD  
52 motif that is formed by a conserved His, Lys and Asp in a typical HxKx<sub>4</sub>D sequence that was  
53 recently refined [9]. In plants and mammals, the HKD motif is duplicated in the primary structure,  
54 and both motifs are thought to be closed in the tertiary structure to form the catalytic site at the  
55 interface of the bi-lobed enzyme [10–13].

56 Furthermore, one common mechanism of action defined as a two-step “ping-pong” reaction  
57 mechanism is conserved between plant and mammalian PLDs [2]. The interest in PLD as a drug  
58 target is due to the fact that PLDs are essential to intracellular and extracellular signaling. PLD

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4 59 have been described to be involved in settings ranging from platelet activation to response to  
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7 60 cardiac ischemia, viral infections, neurodegenerative diseases, and in cancers. Therefore, human  
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9 61 PLD are a valid target for a variety of drug therapies [14].

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11 62 Our work focuses on identifying potential new inhibitor of eukaryotic PLDs, especially the human  
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14 63 isoforms, by using a plant PLD as a model. Indeed, VuPLD shows an overall 19 % peptidic  
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16 64 sequence identity with human PLD1, which reaches 42 % and 69 % around the first and second  
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19 65 HKD motif, respectively. Thus, based on these elements, we hypothesized that an inhibitor  
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21 66 directed against the catalytic site of plant PLD would somehow be able to inhibit mammalian PLD  
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24 67 and *vice versa*. Previous works to screen inhibitors on mammalian PLD were weakened by the  
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26 68 difficulty to obtain sufficient amounts of enzyme for HTS of PLD inhibitors. This difficulty was  
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29 69 partially circumvented, either by the use of the atypical prokaryotic PLD enzyme from  
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31 70 *Streptomyces* for preliminary screenings [15], by using *in silico* methods [16], by virtual screening  
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34 71 [17], or other undisclosed conditions [18]. Recombinant plant PLD $\alpha$  is however easy to produce  
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36 72 and could be purified in one step [19]. Therefore, considering their similarities at multiple levels,  
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39 73 we assumed that a molecule capable of inhibiting plant PLDs could as well affect human PLDs.  
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41 74 This strategy could thus constitute an interesting and original approach to identify novel inhibitors  
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43 75 than using non-HKD PLDs as templates for the screening.

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45 76 Only a few inhibitors of PLDs were described in the literature. N-acylethanolamines (NAEs) are  
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48 77 a class of potent lipids released from the membrane phospholipid N-acylphosphatidylethanolamine  
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51 78 (NAPE), and are found as endogenous constituents of desiccated seeds. The hydrolysis of NAPE  
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53 79 is catalyzed by a NAPE-specific PLD or PLD-like enzyme [20], but not by canonical PLD such as  
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56 80 plant PLD $\alpha$ . However, NAE is able to inhibit PLD $\alpha$  activity with an IC<sub>50</sub> estimated at 0.15  $\mu$ M as  
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58 81 for NAE (12:0) [21]. Applying NAE (12:0) improved seed desiccation tolerance via the inhibition  
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82 of PLD $\alpha$ , that led to the reduction of PA formation, PA being a key modulator of desiccation  
83 sensitivity [22]. Likewise, lysophosphatidylethanolamine (LPE) was shown to be a specific  
84 inhibitor of plant PLDs, that could act as a lipid-derived senescence retardant via the inhibition of  
85 PLD [23]. As an example adding 10  $\mu$ M of LPE (18:1) reduced by half the activity of PLD partially  
86 purified from cabbage [23].

87 Similarly, the immunosuppressant ciclosporin at 5  $\mu$ M was also shown to completely inhibit the  
88 cabbage PLD activity [24], and so were alkylphosphate esters [25], octadecylphosphocholine  
89 being the most efficient inhibitor of this class of molecules with an IC<sub>50</sub> of 6.4  $\mu$ M. However, these  
90 molecules having a classical phosphoester bond they are substrate as well of PLD enzymes [26].  
91 In fact, compared to PC, the canonical substrate tested in the same conditions, around 3% of the  
92 specific activity was found when hexadecylphosphocholine was used [25]. Hexanal and its reduced  
93 form hexanol have been shown to be effective PLD inhibitors in different subcellular fractions  
94 from corn kernels, and PLD activity was inhibited from 50 to 70 % using 0.1 % of hexanal in *in*  
95 *vitro* assays [27,28]. Volatile hexanal being a natural product of the catabolism of linoleic and  
96 linolenic acid during senescence, it was used as a check-point for arresting membrane lipid  
97 degradation in plants, and consequently enhances shelf-life of fruits as described in raspberry [29]  
98 and in litchi [30]. PLD inhibition is used in banana when treated with hexanal, to thicken the cell  
99 wall and impeded penetration of pathogenic spores [31]. Moreover, PLD in rice causes rancidity and  
100 'stale flavor' in bran oil, and thus limits the rice bran usage [32], demonstrating the need for specific  
101 plant PLD inhibitors.

102 As far as mammalian PLDs are concerned, several compounds were reported in literature. For  
103 instance, fodrin [33] indirectly inhibits the PLD activity by decreasing the amount of  
104 phosphatidylinositol (4,5)-bisphosphate (PIP<sub>2</sub>), a cofactor necessary for mammalian PLD activity.



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4 105 However, this compound is not considered useful for research or clinical trials. Halopemide was  
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6 106 identified by HTS in 2007 as a modest inhibitor of PLD2 [18]. Several variants of halopemide  
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9 107 were then developed to exhibit a preferential selectivity towards either PLD1 or PLD2 [34]. *In*  
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11 108 *vitro*, halopemide has an IC<sub>50</sub> of 220 and 310 nM for PLD1 and PLD2 respectively. Conversely,  
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14 109 VU0155069 is more selective towards PLD1, with an IC<sub>50</sub> of 46 nM towards PLD1 compared to  
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16 110 933 nM towards PLD2 *in vitro*, and 11 nM and 1800 nM in cells respectively, whereas a  
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19 111 VU0364739 precursor is more selective towards PLD2 (IC<sub>50</sub> of 140 nM *in vitro* and of 110 nM in  
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21 112 cells, compared to 5.1 μM *in vitro* and 1 μM in cells towards PLD1) [35]. Interestingly 5-fluoro-  
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24 113 2-indolyl des-chlorohalopemide (FIPI) which at approximately 25 nM induced a 50% loss of  
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26 114 activity independently on PLD1 and PLD2 [36] is thought to act on the catalytic site of both  
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29 115 isoforms PLD1 and PLD2. In the opposite VU0364739 which is selective of PLD2 [37] was  
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31 116 previously shown by computational modelling to use an allosteric site overlapping with the PH  
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33 117 domain to target its activity [38], but surface plasmon resonance experiments have demonstrated  
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36 118 that this family of inhibitors binds directly to the catalytic domain [11]. FIPI's IC<sub>50</sub> on PLD from  
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38 119 *Streptomyces* strain PMF was reported to be around 100 nM, which is 4 to 6 times higher than the  
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41 120 values found on PLD1 and PLD2, demonstrating that some inhibitors directly block enzyme and  
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43 121 phospholipid vesicles substrate binding [15]. Last but not least, resveratrol, an antioxidant found  
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46 122 in red wine, was described to inhibit plant PLD [39], and human isoforms as well with an IC<sub>50</sub>  
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48 123 around 61 μM [40].  
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50 124 In order to find the optimal conditions for the screening and to understand the best features of a  
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53 125 PLD inhibitor, we first characterized the purified enzyme VuPLD and described its physical shape  
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55 126 and biochemical structural properties. Then, we characterized the effect of already-known  
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58 127 inhibitors of eukaryotic PLD on VuPLD. Several types of inhibitors have been described in the  
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4 128 literature and could be basically divided into three classes. First, phosphate mimicking ions such  
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6 129 as vanadate [41] or tungstate that compete with the phosphate moiety of the phospholipid substrate.  
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9 130 Secondly natural products such as resveratrol [39] and ciclosporine [24], that are small organic  
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11 131 compounds supposedly protecting enterocyte mitochondria from oxidative stress. Finally,  
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14 132 halopemide and its derivatives that were specially designed for the two isoforms of mammalian  
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16 133 PLD with catalytic activity (PLD1 and PLD2). We then screened a chemical library of roughly  
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19 134 3,120 molecules of diverse origins. The optimization of a 6-step screen could now be used for  
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21 135 larger libraries.  
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## 25 26 137 **Material and Methods**

### 27 28 29 138 **Reagents and material**

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33 139 1,2-Dimyristoyl-*sn*-glycero-3-phosphocholine (DMPC), choline oxidase from *Arthrobacter*  
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35 140 *globiformis*, horseradish peroxidase (type VI), bovine serum albumin (BSA), 8-hydroxyquinoline  
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38 141 (8HQ), resveratrol, ciclosporin, VU0285655, alcohols, hexanal, N-acylethanolamine, and mineral  
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40 142 salts: sodium vanadate, vanadate derivatives (oxytriethoxide, oxytripropoxide, oxytriisopropoxide,  
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43 143 tris(triphenylsiloxy) ), tungstate and phosphate came from Sigma Aldrich-Fluka Chimie.  
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45 144 Halopemide, VU0359595, and FIPI were from Cayman Chemical. Palmitoyl-2-oleoyl-*sn*-glycero-3-  
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48 145 phosphocholine (POPC) and 1-palmitoyl-2-oleoyl-*sn*-glycero-3-phosphate (POPA) in chloroform  
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50 146 were purchased from Avanti Polar Lipids (Alabama, USA). Thin layer chromatography (TLC)  
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53 147 plates were purchased from Macherey-Nagel GmbH & Co. Kg (Germany). Mammalian-PLDs  
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55 148 inhibitors halopemide and derivatives were from Cayman chemicals. Mineral salts: sodium  
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58 149 vanadate and its derivatives, tungstate and phosphate were dissolved in water. All PLD inhibitors  
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60 150 and compounds from the ICBMS chemical library were prepared in DMSO (DMSO levels used  
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151 in this study remained below 5%). Flat-bottom black 96-well microplates (Greiner bio-one) were  
152 used for fluorescence measurements.

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**154 Purification of VuPLD**  
155 Recombinant VuPLD was produced in *Pichia pastoris* and purified as described previously [19].  
156 Small aliquots were thawed and diluted before each experiment. A single batch was used for the  
157 enzymatic characterization, and another for the screening of the chemical library. Protein  
158 concentrations were determined using the Bradford's procedure [42], with Dye Reagent and BSA  
159 as the standard.

160  
**161 Small angle X-ray scattering experiment**  
162 Data were collected at the ESRF beamline BM29 (Grenoble, France). VuPLD was concentrated  
163 at 2.55 mg/mL in PIPES buffer 10 mM, pH 6.2, containing EDTA 0.1 mM and NaCl 200 mM.  
164 Reconstruction of the overall shape of the VuPLD from X-ray scattering data was first achieved  
165 by the *ab initio* modeling programs DAMMIN (20 models calculated) [43] or GASBOR (50  
166 models calculated) [44], then models were averaged using DAMAVER [45]. Comparison of the  
167 SAXS data and protein structure (PDB code : 6KZ9) was done using FoXS [46]. Protein molecular  
168 mass was estimated from scattering intensities  $I(0)$  using SAXSMoW2 software [47].

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**170 PLD activity measurement**

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4 171 *Chromogenic assay*

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7 172 A detailed protocol of this assay has been recently published [48]. Briefly, 0.4 mM of DMPC in  
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10 173 SDS/Triton-X100 mixed micelles were incubated with pure PLD (0.7 to 0.9  $\mu\text{g}/\text{mL}$ , final  
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12 174 concentration) in the presence or absence of inhibitors for 10 min at 30°C. The reaction was started  
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15 175 by the addition of 20 mM of  $\text{CaCl}_2$ , then blocked after 10 min by addition of 83.3 mM of EDTA,  
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17 176 and revealed by addition of peroxidase, choline oxidase, 2-hydroxy-3,5-dichlorobenzenesulfonate  
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20 177 and 4-aminoantipyrine. The PLD-generated choline was quantified with end point measurements  
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22 178 by recording the absorbance at 500 nm, based on a standard curve obtained with pure choline.  
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29 180 *Fluorescence-based assay*

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31 181 The preparation of 8HQ / phospholipid/ detergent mixtures was adapted from [49] with minor  
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33 182 modifications to control the activation of the enzymes. To prepare the mixtures, 40 nmol of POPC  
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35 183 (266  $\mu\text{M}$ , final concentration) were first dissolved in chloroform, then evaporated under a stream  
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38 184 of nitrogen. Subsequently, dried phospholipids were dispersed in a buffer containing 50 mM Tris-  
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41 185 HCl, pH 8.0, 1 mM Triton X-100, 0.7 mM SDS, 66  $\mu\text{M}$  8HQ, and 50  $\mu\text{M}$  EDTA. The micellar  
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43 186 solution was vortexed for 30 s, sonicated for 10 min using a bath sonicator (Deltasonic type O11C,  
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46 187 Fécamp, France), then vortexed for an additional 30 s. One pmol of enzyme (VuPLD) was finally  
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48 188 added as well as the compounds to be tested. After a 10-minute incubation period at 30°C, the  
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51 189 reaction was triggered by adding 20 mM of  $\text{CaCl}_2$ . Fluorescence measurements were recorded at  
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53 190 30°C using a microtiter plate fluorescence reader (Tecan Infinite M200, Austria) at wavelengths  
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55 191 366 nm and 490 nm of excitation and emission, respectively. To monitor PLD activity, a baseline  
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58 192 of 4 kinetic cycles of 30 s was first recorded, after which 20 mM  $\text{CaCl}_2$  were injected in each well  
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193 to trigger the PLD hydrolysis. Fluorescence intensities were then measured during a 1h-kinetic  
194 cycle with interval shaking before each read. The PLD-generated PA was quantified by recording  
195 the fluorescence in the same conditions as described above, based on a standard curve obtained  
196 with pure POPA [49].

197  
198 **TLC analysis**

199 The phospholipids were extracted with chloroform/methanol (2:1, v/v). The lower organic phase  
200 was collected, dried, and subjected to TLC separation together with proper phospholipid standards.  
201 TLC plates were developed in a solvent system using a mixture of chloroform/methanol/water  
202 (65:25:5, v/v/v) as the eluent. Phospholipid spots were revealed by charring the plate after spraying  
203 a mixture of 10 % copper sulfate and 10 % phosphoric acid in water and heating at 150°C for 5  
204 min.

206 **Statistical analysis**

207 Statistically significant differences were determined by Brown-Forsyth and Welsh ANOVA test  
208 using the computer program Prism (GraphPad Software). Results were considered significant  
209 when  $p < 0.05$  (\*), highly significant when  $p < 0.01$  (\*\*), and extremely significant when  $p < 0.001$   
210 (\*\*\*).

212 **Results**

213 **Modelization of VuPLD by SAXS experiment**

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4 214 VuPLD was produced according to Arhab *et al.*[48]. The amount of homogenous VuPLD obtained  
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6 215 after purification was 1 mg per liter of yeast culture. The specific activity toward DMPC was 20  
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9 216 U/mg. Scattering measurement with synchrotron radiation were performed at pH 6.2 with 2.55  
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11 217 mg/mL of VuPLD. A typical SAXS profile of VuPLD is shown in Fig. 1A and a Kratky curve  
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14 218 displays a symmetric peak indicating that VuPLD is globular and correctly folded (Fig. 1B). From  
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16 219 scattering intensities  $I(0)$  a molecular mass of 89.1 kDa was calculated, which is in accordance  
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19 220 with a monomer of VuPLD as the theoretical molecular mass deduced from the amino acid  
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21 221 sequence is 91.5 kDa. The radius of gyration is 2.81 +/- 0.05 nm and  $D_{max}$  (maximum dimension)  
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24 222 is calculated to be 9.89 nm (Fig. 1C). The calculated model by DAMMIN shows a globular and  
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26 223 symmetric molecule with extended extremities and an ovoid shape (Fig. 1D). The program  
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29 224 GASBOR calculated a very similar model however this one is less symmetrical and displays a  
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31 225 larger base and a fine apical extremity (Fig. S1). The comparison of the collected SAXS data and  
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33 226 calculated scattering curve from the published structure of the PLD show a good correlation (Fig.  
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36 227 S2A) indicating that the protein adopts a similar conformation in the crystals and in solution. The  
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38 228 small differences observed could be due to small motion of the C2 domain.  
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#### 42 43 230 **Characterization of the salts effect on PLD activity**

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45 231 Ions such phosphate, tungstate and vanadate salts were already reported as potent inhibitors of  
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48 232 partially purified plant PLDs. We were able to confirm these results by measuring an  $IC_{50}$  for these  
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50 233 three ions on a pure VuPLD. Vanadate was the most potent of them with an  $IC_{50}$  of 58  $\mu$ M and  
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53 234 being mostly able to totally inhibit the enzymatic activity at a concentration of 260  $\mu$ M (Fig. 2A)  
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55 235 and was consequently used as a reference for HTS of PLD inhibitors. Likewise, tungstate was also  
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58 236 able to inhibit totally the PLD activity at a concentration of 2 mM, and harbored an  $IC_{50}$  value of  
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4 237 375  $\mu\text{M}$  (Fig. 2C). Phosphate behaved in between, with an  $\text{IC}_{50}$  estimated to 125  $\mu\text{M}$  but not being  
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7 238 able to totally abolish the activity, 25% of residual activity was quantified at 1 mM of phosphate  
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9 239 that is the maximal concentration usable in this test (Fig. 2B), as  $\text{Ca}^{2+}$  cofactor precipitates at higher  
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12 240 concentration of phosphate.

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14 241 An unusual slight synergistic and inhibitory effect was observed when phosphate and vanadate  
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16 242 were incubated together (Fig. 2D). Indeed, when both molecules were incubated together at their  
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19 243 respective  $\text{IC}_{50}$ , only 15 % of residual activity was measured that was lower than the theoretical  
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21 244 25 % expected at these particular concentrations, suggesting the combined salts potentiate the PLD  
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24 245 inhibition.

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#### 27 28 29 247 **Vanadate derived compounds inhibitory effect on PLD activity**

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31 248 Vanadate being the PLD inhibitor with the more potent inhibitory impacts, we decided to examine  
32  
33 249 the effects of the environment around this particular ion by testing diverse vanadate derivatives  
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36 250 such as vanadium oxytriethoxide, vanadium oxytripropoxide, vanadium oxytriisopropoxide and  
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38 251 tris(triphenylsiloxy) vanadium oxide, as all these molecules harbor different hydrocarbon chain  
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41 252 lengths and steric environment (Fig. S3). If vanadate took the place of the phosphate moiety of  
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43 253 phospholipid substrate in the active site, changes of alkyl chain length and steric environment  
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46 254 around vanadate would have been expected to modify its  $\text{IC}_{50}$ . However, even if the  $\text{IC}_{50}$  of  
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48 255 vanadate derivatives were lower than the one of vanadate (Fig. S4), no dramatic variation could  
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51 256 be observed amongst these four derivatives (Fig. 3).  $\text{IC}_{50}$  were ranging from 7  $\mu\text{M}$  (vanadate  
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53 257 oxytriethoxide) to 15  $\mu\text{M}$  (vanadate oxytriisopropoxide).

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58 259 **Alcohols inhibitory effect on PLD hydrolysis activity**

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Historically, primary alcohols have been used as inhibitors of PLD hydrolysis activity. However, one must state that it is not a proper inhibitory effect as the use of alcohol derives PLD activity from hydrolysis *stricto sensu* toward the transphosphatidylation activity. In fact, alcohols can be used as secondary substrate and therefore PLD can transfer the phosphatidyl group to a primary alcohol to generate new phospholipids. The use of butan-1-ol is quite common to follow mammalian PLD activity by detecting the production of phosphatidylbutanol. We consequently tried to define which primary alcohol would be the best to avoid PLD-generated PA by testing the primary alcohols from methanol to hexan-1-ol (Fig. 4).

As expected, methanol and ethanol were not efficient secondary substrates of VuPLD. For short-chain alcohol the best candidate was butan-1-ol with an  $IC_{50}$  of 150 mM compared to propan-1-ol which was more than twice less efficient with 320 mM. Pentan-1-ol had an  $IC_{50}$  of 60 mM which is the best among all the alcohols tested. Finally, hexan-1-ol seemed not to be a proper secondary substrate as the activity quickly falls at 20 mM probably indicating enzyme denaturation (Fig. 4A). All secondary alcohols tested appeared not to be proper substrates as they displayed high  $IC_{50}$ , 160 mM for pentan-2-ol, 360 mM for butan-2-ol and propan-2-ol had no effect on PLD activity (Fig. 4A).

However, these values are based on the fluorimetric detection of PA (see Material & methods section). A more convenient way to detect phosphatidylalcohols issued from the PLD-catalyzed transphosphatidylation reaction is to use TLC for phospholipids separation. As can be seen from Fig. 4B, from all alcohols tested, the transphosphatidylation products were qualitatively detected only with methanol, ethanol, propan-1-ol and butan-1-ol.

#### **Effect of known mammalian PLD inhibitors on VuPLD**



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4 283 Halopemide was shown previously to inhibit both human PLD1 and PLD2 isoforms, VU0359595  
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6 284 targets the PLD1 isoform with an  $IC_{50}$  calculated to be 3.7 nM, VU0285655 targets the PLD2  
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9 285 isoform with an  $IC_{50}$  calculated to be 635 nM. When tested on VuPLD, no inhibition could be  
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12 286 observed up to 10  $\mu$ M (Fig. 5). The PLD activity was reduced down to 50 % when 100  $\mu$ M of each  
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14 287 these three inhibitors were used independently.

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### 19 289 **Test of alleged plant PLD inhibitors**

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21 290 Amongst the molecules reported in literature to inhibit plant PLD activity we tested both  
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24 291 ciclosporin A and resveratrol on VuPLD using our fluorescence-based assay. In our hand, neither  
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26 292 ciclosporin A (Fig. 6A) nor resveratrol (Fig. 6B) were able to significantly affect VuPLD activity.  
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29 293 A slight 20% PLD inhibition could be observed using 2 mM of resveratrol, which is far from the  
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31 294 value reported by others where more than 90% of inhibition was measured with 130  $\mu$ M of  
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33 295 resveratrol [39]. As far as ciclosporin A is concerned, no inhibition was detected until 100  $\mu$ M  
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36 296 even if a quasi-total inhibition was previously reported for the PLD from cabbage using 5  $\mu$ M of  
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38 297 this compound [24,39].

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41 298 Similarly, neither NAE (Fig. 6C) nor hexanal (Fig. 6D) which have been demonstrated as efficient  
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43 299 inhibitors of castor bean [21] or raspberry [29] PLDs, respectively, displayed any inhibitory effect  
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45 300 on VuPLD.

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### 50 302 **Screening of new PLD inhibitors**

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53 303 Because of its sensitivity and ease of use, we aimed to validate the 8HQ-based assay suitability for  
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55 304 HTS of novel PLD inhibitors as a start. In order to do so, we calculated the  $Z'$ -factor, a statistical  
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58 305 parameter used to evaluate the quality and performance of an assay. This parameter is reflective

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4 306 of both the assay signal dynamic range and the data variation associated with signal measurements  
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6 307 [50]. In light of former results showing sodium vanadate as the most potent inhibitor of VuPLD  
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9 308 (Fig. 2B), we consequently decided to use it as a reference and a positive control for the screening.  
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11 309 Briefly, confronting the variability of 30 values of PLD activity measured with either 100  $\mu$ M of  
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14 310 vanadate or just 3 % DMSO, we obtained a calculated value of 0.803 for the  $Z'$ -score (Fig. 7A).  
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16 311 This value being comprised in the in the interval between 0.5 and 1.0 thus qualifies the 8HQ-based  
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19 312 test as “excellent” [50] for HTS of PLD inhibitors.  
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21 313 Moreover, as most of chemical library are DMSO solubilized, DMSO was shown only to have a  
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24 314 slight effect on PLD activity up to 5 % DMSO final concentration, but no statistical difference  
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26 315 compared from the control was observed when 5% DMSO final concentration was used (data not  
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29 316 shown).  
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31 317 We built a six-step protocol for the HTS of a PLD inhibitor (Fig. 7B). We consequently performed  
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33 318 a small-scale screening on the chemical library of our institute that is composed of 3,120  
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36 319 compounds from diverse origins. These molecules include quinolones, indoles, indolinones,  
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38 320 benzofurans, coumarins and flavonoids from chemistry and pharmaceutical laboratories.  
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41 321 A hit was considered every molecule capable to activate or inhibit PLD activity *in vitro* by more  
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43 322 than 50%. Out of 3,120 initial compounds, 227 molecules were found to reduce the PLD activity  
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46 323 and 53 to activate this activity (the others were found to have little or no effect at all). Because  
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48 324 some of the hits could be fluorescent molecules, or molecules with putative screen effect on the  
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51 325 8HQ probe, or molecules that could chelate the calcium ions for example, *id est* molecules that  
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53 326 could interfere with the fluorescent assay, we screened the putative 277 inhibitory molecules using  
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56 327 chromogenic assay which quantifies the free choline released by the PLD. After this step, 11  
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58 328 molecules were retained then further checked for their absence of effect on a PA standard curve  
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329 fluorescence. Finally, the 2 remaining compounds were tested on crude extract of *Pichia pastoris*  
330 expressing VuPLD to avoid unspecific protein interaction. The last hit was confirmed against other  
331 HKD-type PLDs like cabbage PLD [51] or peanut PLD [52]. Then, it was excluded after the last  
332 step of this protocol as *de novo* synthesis of the molecule showed no effect at all and as several  
333 contaminants were present in the initial batch of molecule used to make up the library.

## 335 Discussion

336 Using two different *ab initio* modelization programs SAXS data indicate that VuPLD is a globular  
337 enzyme with elongated tops, thus the enzyme seems overall symmetrical with an ovoid shape.  
338 These models do not allow the positioning of the regulatory C2 domain. The determination of the  
339 overall shape of VuPLD is in accordance with the results obtained with cabbage PLD $\alpha$ 2 in 2012  
340 by others [53]. Both models of nearly the same length (809 residues for VuPLD compare to 812  
341 for cabbage PLD $\alpha$ 2) share 81.3% amino acid sequence identity and have a similar gyration radius,  
342 but the  $D_{max}$  is lower in cabbage PLD (2.81 +/- 0.05 nm (this study) vs 2.89 +/- 0.01 nm [53])  
343 indicating a slightly smaller enzyme. Also, the porod volume of VuPLD is more important  
344 indicating a larger enzyme of 140.8 nm<sup>3</sup> vs 126.7 nm<sup>3</sup> for cabbage PLD [53]. These very slight  
345 differences for such similar enzymes in terms of sequence might be explained by the fact that, even  
346 if they are purified the same way, they are not recombinantly expressed in the same organism.  
347 Specifically, cabbage PLD was expressed in *Escherichia coli* BL21 (DE3), contrary to VuPLD  
348 that was expressed in yeast *Pichia pastoris* X33 cells which are known to glycosylate proteins.  
349 The structure of PLD $\alpha$ 1 from Arabidopsis that was recently solved [12] positioned the C2 domain  
350 as a small protuberance interacting with the second HKD domain which is, however, not in  
351 accordance with the ovoid shape with extended extremities of the VuPLD obtained in this work.

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4 352 Nevertheless, the SAXS model of VuPLD (this work) and of PLD from cabbage [53], or crystal  
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6 353 structure of *Arabidopsis* PLD [12] are very close (Fig. S2B) and interchangeable, which makes of  
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9 354 VuPLD another PLD model, and specially a convenient tool to study PLD inhibition as this latest  
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12 355 protein could be recombinantly expressed and easily purified in one step [54].  
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14 356 To prove that VuPLD is a relevant model to study plant PLD inhibition, we started with the use of  
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16 357 substrate analogs, in particular, analogs of the phosphate head of phospholipid. Therefore, we  
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19 358 tested phosphate salt at first sight and structurally close chemicals like vanadate or tungstate. These  
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21 359 salts are known to modulate PLD activity and we show here that vanadate is the best of them with  
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24 360 the finest IC<sub>50</sub>. Consequently, we tested vanadate derivatives that differ by their hydrophobic  
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26 361 properties. Indeed, vanadate and then vanadate oxytriethoxide, vanadate oxytripropoxide,  
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29 362 vanadate oxytriisopropoxide and tris(triphenylsiloxy) vanadium oxide are increasing hydrophobic  
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31 363 molecules. Among them, vanadate oxytriethoxide that only display ethane moieties around  
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34 364 vanadate salt is the best inhibitor. These results may suggest that the binding site of vanadate on  
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36 365 PLD was not influenced by the alkyl chain length and/or steric environment and has enough room  
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38 366 to allow the fixation of the different derivatives tested.  
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41 367 Another way to inhibit the hydrolase activity of PLDs is the use of alcohols, keeping in mind that  
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43 368 alcohols are not proper inhibitors but are just another secondary substrate competing with water to  
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46 369 induce a transphosphatidylolation reaction. Here, we show that short-chained alcohols like methanol,  
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48 370 ethanol and to a lesser extent propan-1-ol do not affect the hydrolysis activity. Secondary alcohols  
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51 371 are also not substrates. The commonly used butan-1-ol is definitely the best primary alcohol  
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53 372 susceptible to generate a phosphatidylbutanol at a weak concentration. With other alcohols, no  
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55 373 transphosphatidylolation products were visible on TLC plates, meaning that they are not proper  
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4 374 secondary substrates of PLD. But one has to notice the fact that they indeed reduce PLD activity.  
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7 375 As stated above for hexan-1-ol, propan-2-ol and pentan-1-ol probably induce enzyme denaturation.  
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9 376 Human PLD inhibitors do not show any inhibitory effect on plant PLD, even if these PLD [11]  
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11 377 [12] share a common and superposable active site (Fig. S5). These results ask questions about the  
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14 378 mechanism by which they inhibit human PLDs. We demonstrated recently that all PLDs share  
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16 379 common sequence motifs [9]. Among them a pocket constituted by two HKD motifs form the  
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19 380 catalytic site. But also, several similar sequences are found in all eukaryotic PLDs. Indeed, PLDs  
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21 381 are mostly differing by their regulatory motifs [9]. For example, human PLD bears a PX and PH  
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24 382 domain [2] whereas plant PLD bears a C2-domain responsible for the fixation of  $Ca^{2+}$  ions, a  
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26 383 mandatory activator of plant PLD [55]. However, the catalytic sites of *Arabidopsis* PLD [12] and  
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29 384 human PLD1 [13] share a common feature as globular fold with a funnel-shaped cavity leading to  
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31 385 the active site. Considering the localized relative sequence similarity between human PLD and  
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33 386 plant PLD, and considering that if human PLD inhibitors were targeting the catalytic site they  
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36 387 would have a similar effect on plant PLD, it is possible that human PLD inhibitors are targeting  
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38 388 (i) aminoacid sequences that do not exist in plant PLD or (ii) the regulatory domains and therefore  
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41 389 do not inhibit the catalytic activity of the enzyme but its capacity to bind its partners and cofactors.  
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43 390 The structure of the catalytic domains of human PLD1 and PLD2 that was recently published [11]  
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46 391 would surely help to explain the selectivity of several molecules thought or used as generic  
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48 392 inhibitors on plant and/or mammalian PLDs. As it was not been possible to solve the structure of  
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51 393 PLD $\alpha$ 1 with FIPI [12], this compound is assumed to localized in the substrate-binding pocket of  
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53 394 the enzyme. However, our direct and continuous fluorescence based assay failed to display an  
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55 395 inhibitory effect on VuPLD, as no inhibition could be observed up to 100  $\mu$ M (Fig. 5) and only an  
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58 396 apparent 29% reduction of activity was observed when 1 mM of FIPI was used (data not shown),  
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4 397 noting that such a high concentration would probably interfere with the fluorescence of the 8HQ  
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6 398 probe. These differences of FIPI effects could be explained by the nature of the plant PLDs used,  
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9 399 as some residues are dissimilar in this pocket among the plant PLDs, or by the nature of the test,  
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11 400 the activity being assayed in a direct and continuous manner in this work.  
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14 401 At the exception of vanadate that is still the more potent plant PLD inhibitor to our knowledge,  
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16 402 almost of all plant PLD inhibitors described in the literature failed to inhibit VuPLD, consequently  
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19 403 there is still a need for a specific plant PLD inhibitor or a generalist eukaryotic PLD inhibitors  
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21 404 acting both on plant and mammalian PLDs. Looking closely, it appears that the use of a purified  
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24 405 enzyme in this study is the major difference compared to all other papers published on this topic.  
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26 406 In this regard, it is possible that these compounds interfere with others enzymes used in the cascade  
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29 407 of choline quantification. On the other hand, in our case the use of an assay to directly measure the  
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31 408 PA released directly from PLD-catalysed hydrolysis, constitutes a significant advantage to  
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33 409 quantify PLD activity [49].  
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36 410 Indeed, this 8HQ-based assay was statistically validated with a  $Z'$  score of 0.803 indicating an  
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38 411 excellent test for HTS and demonstrating that vanadate should be used as a positive control for  
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41 412 forthcoming screenings. We consequently screened as a proof of concept a chemical library of  
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43 413 ~3,120 compounds. After six runs of enzymatic tests combining successively the advantages of  
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46 414 both fluorescent and chromogenic assays to avoid false positives, no hit could unfortunately be  
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48 415 retained as the unique potential candidate appeared to be a contaminant. In the future, combining  
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51 416 the two PLD enzymatic assays with their specific strengths and weaknesses, their distinct and  
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53 417 alternative properties, larger libraries could be screened to identify and validate new PLD  
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55 418 inhibitors.  
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4 420 In conclusion, this work highlights (i) the necessity for a novel universal inhibitor potentially  
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6 421 targeting the catalytic site of PLDs, (ii) the potential of a purified plant PLD to screen PLD  
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9 422 inhibitors and (iii) the need to screen larger chemical libraries to find new inhibitory molecules.  
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36 432 **Author Contributions**

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38  
39 433 YA and MVC performed SAXS and modeling experiments. AC and FP contributed to compound  
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41 434 design and synthesis. YA, KB, HA, MA, RR performed biochemical assays. LB, SM, AA and AN  
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43  
44 435 devised the project and the main conceptual ideas. AN was in charge of funding acquisition. All  
45  
46 436 authors contributed to data interpretation. The manuscript was written from contributions by all  
47  
48  
49 437 the authors. All the authors have given approval to the final version of the manuscript.  
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55 439 **Competing interests**

56  
57  
58 440 The authors have no competing interests to declare.  
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## Abbreviations

8-Hydroxyquinoline (8HQ), high-throughput screening (HTS), lysophosphatidylethanolamine (LPE), N-acylethanolamines (NAE), N-acylphosphatidylethanolamine (NAPE), phosphatidic acid (PA), phospholipase D (PLD), palmitoyl-2-oleoyl-*sn*-glycero-3-phosphocholine (POPC).

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## References

- [1] S.M. Hammond, Y.M. Altshuller, T.C. Sung, S.A. Rudge, K. Rose, J. Engebrecht, A.J. Morris, M.A. Frohman, Human ADP-ribosylation factor-activated phosphatidylcholine-specific phospholipase D defines a new and highly conserved gene family, *J. Biol. Chem.* 270 (1995) 29640–29643.
- [2] P.E. Selvy, R.R. Lavieri, C.W. Lindsley, H.A. Brown, Phospholipase D: enzymology, functionality, and chemical modulation, *Chem. Rev.* 111 (2011) 6064–6119. <https://doi.org/10.1021/cr200296t>.



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4 463 [3] A. Marchler-Bauer, Y. Bo, L. Han, J. He, C.J. Lanczycki, S. Lu, F. Chitsaz, M.K. Derbyshire,  
5 464 R.C. Geer, N.R. Gonzales, M. Gwadz, D.I. Hurwitz, F. Lu, G.H. Marchler, J.S. Song, N.  
6 465 Thanki, Z. Wang, R.A. Yamashita, D. Zhang, C. Zheng, L.Y. Geer, S.H. Bryant,  
7 466 CDD/SPARCLE: functional classification of proteins via subfamily domain architectures,  
8 467 *Nucleic Acids Res.* 45 (2017) D200–D203. <https://doi.org/10.1093/nar/gkw1129>.  
9  
10 468 [4] S. Imamura, Y. Horiuti, Purification of *Streptomyces chromofuscus* phospholipase D by  
11 469 hydrophobic affinity chromatography on palmitoyl cellulose, *J. Biochem. (Tokyo)*. 85 (1979)  
12 470 79–95.  
13  
14 471 [5] S. Lee, K.R. Lynch, Brown recluse spider (*Loxosceles reclusa*) venom phospholipase D  
15 472 (PLD) generates lysophosphatidic acid (LPA), *Biochem. J.* 391 (2005) 317–323.  
16 473 <https://doi.org/10.1042/BJ20050043>.  
17  
18 474 [6] S.A. Rudge, J. Engebrecht, Regulation and function of PLDs in yeast, *Biochim. Biophys.*  
19 475 *Acta.* 1439 (1999) 167–174.  
20  
21 476 [7] K. Rose, S.A. Rudge, M.A. Frohman, A.J. Morris, J. Engebrecht, Phospholipase D signaling  
22 477 is essential for meiosis, *Proc. Natl. Acad. Sci. U. S. A.* 92 (1995) 12151–12155.  
23  
24 478 [8] I. Leiros, F. Secundo, C. Zambonelli, S. Servi, E. Hough, The first crystal structure of a  
25 479 phospholipase D, *Struct. Lond. Engl.* 1993. 8 (2000) 655–667.  
26  
27 480 [9] Y. Arhab, A. Abousalham, A. Noiriél, Plant phospholipase D mining unravels new conserved  
28 481 residues important for catalytic activity, *Biochim. Biophys. Acta Mol. Cell Biol. Lipids.* 1864  
29 482 (2019) 688–703. <https://doi.org/10.1016/j.bbali.2019.01.008>.  
30  
31 483 [10] C.P. Ponting, I.D. Kerr, A novel family of phospholipase D homologues that includes  
32 484 phospholipid synthases and putative endonucleases: identification of duplicated repeats and  
33 485 potential active site residues, *Protein Sci. Publ. Protein Soc.* 5 (1996) 914–922.  
34 486 <https://doi.org/10.1002/pro.5560050513>.  
35  
36 487 [11] C.M. Metrick, E.A. Peterson, J.C. Santoro, I.J. Enyedy, P. Murugan, T. Chen, K. Michelsen,  
37 488 M. Cullivan, K.A. Spilker, P.R. Kumar, T.L. May-Dracka, J.V. Chodaparambil, Human PLD  
38 489 structures enable drug design and characterization of isoenzyme selectivity, *Nat. Chem. Biol.*  
39 490 (2020). <https://doi.org/10.1038/s41589-019-0458-4>.  
40  
41 491 [12] J. Li, F. Yu, H. Guo, R. Xiong, W. Zhang, F. He, M. Zhang, P. Zhang, Crystal structure of  
42 492 plant PLD $\alpha$ 1 reveals catalytic and regulatory mechanisms of eukaryotic phospholipase D,  
43 493 *Cell Res.* (2019). <https://doi.org/10.1038/s41422-019-0244-6>.  
44  
45 494 [13] F.Z. Bowling, C.M. Salazar, J.A. Bell, T.S. Huq, M.A. Frohman, M.V. Airola, Crystal  
46 495 structure of human PLD1 provides insight into activation by PI(4,5)P2 and RhoA, *Nat. Chem.*  
47 496 *Biol.* 16 (2020) 400–407. <https://doi.org/10.1038/s41589-020-0499-8>.  
48  
49 497 [14] M.I. McDermott, Y. Wang, M.J.O. Wakelam, V.A. Bankaitis, Mammalian phospholipase D:  
50 498 Function, and therapeutics, *Prog. Lipid Res.* (2019) 101018.  
51 499 <https://doi.org/10.1016/j.plipres.2019.101018>.  
52  
53 500 [15] A. Kulkarni, P. Quang, V. Curry, R. Keyes, W. Zhou, H. Cho, J. Baffoe, B. Török, K.  
54 501 Stieglitz, 1,3-disubstituted-4-aminopyrazolo [3, 4-d] pyrimidines, a new class of potent  
55 502 inhibitors for phospholipase D, *Chem. Biol. Drug Des.* 84 (2014) 270–281.  
56 503 <https://doi.org/10.1111/cbdd.12319>.  
57  
58 504 [16] H. Djakpa, A. Kulkarni, S. Barrows-Murphy, G. Miller, W. Zhou, H. Cho, B. Török, K.  
59 505 Stieglitz, Identifying New Drug Targets for Potent Phospholipase D Inhibitors: Combining  
60 506 Sequence Alignment, Molecular Docking, and Enzyme Activity/Binding Assays, *Chem.*  
61 507 *Biol. Drug Des.* 87 (2016) 714–729. <https://doi.org/10.1111/cbdd.12705>.  
62  
63  
64  
65

- 1  
2  
3  
4 508 [17] D.K. Behera, P.M. Behera, L. Acharya, A. Dixit, Pharmacophore modelling, virtual  
5 509 screening and molecular docking studies on PLD1 inhibitors, SAR QSAR Environ. Res. 28  
6 510 (2017) 991–1009. <https://doi.org/10.1080/1062936X.2017.1393774>.
- 8 511 [18] L. Monovich, B. Mugrage, E. Quadros, K. Toscano, R. Tommasi, S. LaVoie, E. Liu, Z. Du,  
9 512 D. LaSala, W. Boyar, P. Steed, Optimization of halopemide for phospholipase D2 inhibition,  
10 513 Bioorg. Med. Chem. Lett. 17 (2007) 2310–2311. <https://doi.org/10.1016/j.bmcl.2007.01.059>.
- 12 514 [19] Y. Ben Ali, F. Carriere, A. Abousalham, High-level constitutive expression in *Pichia pastoris*  
13 515 and one-step purification of phospholipase D from cowpea (*Vigna unguiculata* L. Walp),  
14 516 Protein Expr Purif. 51 (2007) 162–9. <https://doi.org/10.1016/j.pep.2006.07.018>.
- 15 517 [20] Z. Hussain, T. Uyama, K. Tsuboi, N. Ueda, Mammalian enzymes responsible for the  
16 518 biosynthesis of N-acylethanolamines, Biochim. Biophys. Acta. 1862 (2017) 1546–1561.  
17 519 <https://doi.org/10.1016/j.bbali.2017.08.006>.
- 19 520 [21] S.L. Austin-Brown, K.D. Chapman, Inhibition of phospholipase D alpha by N-  
20 521 acylethanolamines, Plant Physiol. 129 (2002) 1892–1898.  
21 522 <https://doi.org/10.1104/pp.001974>.
- 22 523 [22] H. Chen, X. Yu, X. Zhang, L. Yang, X. Huang, J. Zhang, H.W. Pritchard, W. Li,  
23 524 Phospholipase D $\alpha$ 1-mediated phosphatidic acid change is a key determinant of desiccation-  
24 525 induced viability loss in seeds, Plant Cell Environ. 41 (2018) 50–63.  
25 526 <https://doi.org/10.1111/pce.12925>.
- 27 527 [23] S.B. Ryu, B.H. Karlsson, M. Ozgen, J.P. Palta, Inhibition of phospholipase D by  
28 528 lysophosphatidylethanolamine, a lipid-derived senescence retardant, Proc. Natl. Acad. Sci.  
29 529 U. S. A. 94 (1997) 12717–12721.
- 31 530 [24] M. Madesh, K.A. Balasubramanian, Cyclosporin A inhibits oxidant and calcium stimulated  
32 531 phospholipase D activity in the rat intestinal mitochondria, Biochim. Biophys. Acta. 1389  
33 532 (1998) 206–212.
- 34 533 [25] N. Dittrich, G. Nössner, B. Kutscher, R. Ulbrich-Hofmann, Alkylphosphate esters as  
35 534 inhibitors of phospholipase D, J. Enzym. Inhib. 11 (1996) 67–75.
- 37 535 [26] Y. Ohno, M. Okazaki, I. Hara, Studies on the activity of phospholipase D with alkyl  
38 536 phosphorylcholines, Biochem. Cell Biol. Biochim. Biol. Cell. 68 (1990) 376–379.
- 39 537 [27] G. Paliyath, R.G. Pinhero, R.Y. Yada, D.P. Murr, Effect of processing conditions on  
40 538 phospholipase D activity of corn kernel subcellular fractions, J. Agric. Food Chem. 47 (1999)  
41 539 2579–2588.
- 43 540 [28] K. Tiwari, G. Paliyath, Microarray analysis of ripening-regulated gene expression and its  
44 541 modulation by 1-MCP and hexanal, Plant Physiol. Biochem. PPB. 49 (2011) 329–340.  
45 542 <https://doi.org/10.1016/j.plaphy.2011.01.007>.
- 47 543 [29] W.E. Kayal, G. Paliyath, J.A. Sullivan, J. Subramanian, Phospholipase D inhibition by  
48 544 hexanal is associated with calcium signal transduction events in raspberry, Hortic. Res. 4  
49 545 (2017) 17042. <https://doi.org/10.1038/hortres.2017.42>.
- 50 546 [30] B. Bhushan, S. Kumar, M.K. Mahawar, K. Jalgaonkar, A.S. Dukare, B. Bibwe, V.S. Meena,  
51 547 N. Negi, R.K. Narwal, A. Pal, Nullifying phosphatidic acid effect and controlling  
52 548 phospholipase D associated browning in litchi pericarp through combinatorial application of  
53 549 hexanal and inositol, Sci. Rep. 9 (2019) 2402. <https://doi.org/10.1038/s41598-019-38694-5>.
- 54 550 [31] D. Dhakshinamoorthy, S. Sundaresan, A. Iyadurai, K.S. Subramanian, G.J. Janavi, G.  
55 551 Paliyath, J. Subramanian, Hexanal Vapor Induced Resistance against Major Postharvest  
56 552 Pathogens of Banana (*Musa acuminata* L.), Plant Pathol. J. 36 (2020) 133–147.  
57 553 <https://doi.org/10.5423/PPJ.OA.03.2019.0072>.
- 60  
61  
62  
63  
64  
65

- 1  
2  
3  
4 554 [32] A. Kaur, K. Neelam, K. Kaur, A. Kitazumi, B.G. de Los Reyes, K. Singh, Novel allelic  
5 555 variation in the Phospholipase D alpha1 gene (OsPLD $\alpha$ 1) of wild *Oryza* species implies to  
6 556 its low expression in rice bran, *Sci. Rep.* 10 (2020) 6571. [https://doi.org/10.1038/s41598-](https://doi.org/10.1038/s41598-020-62649-w)  
8 557 [020-62649-w](https://doi.org/10.1038/s41598-020-62649-w).
- 9 558 [33] S. Lukowski, J.P. Mira, A. Zachowski, B. Geny, Fodrin inhibits phospholipases A2, C, and  
10 559 D by decreasing polyphosphoinositide cell content, *Biochem. Biophys. Res. Commun.* 248  
11 560 (1998) 278–284. <https://doi.org/10.1006/bbrc.1998.8942>.
- 12 561 [34] J.A. Lewis, S.A. Scott, R. Lavieri, J.R. Buck, P.E. Selvy, S.L. Stoops, M.D. Armstrong, H.A.  
14 562 Brown, C.W. Lindsley, Design and synthesis of isoform-selective phospholipase D (PLD)  
15 563 inhibitors. Part I: Impact of alternative halogenated privileged structures for PLD1  
16 564 specificity, *Bioorg. Med. Chem. Lett.* 19 (2009) 1916–1920.  
18 565 <https://doi.org/10.1016/j.bmcl.2009.02.057>.
- 19 566 [35] S.A. Scott, P.E. Selvy, J.R. Buck, H.P. Cho, T.L. Criswell, A.L. Thomas, M.D. Armstrong,  
20 567 C.L. Arteaga, C.W. Lindsley, H.A. Brown, Design of isoform-selective phospholipase D  
21 568 inhibitors that modulate cancer cell invasiveness, *Nat. Chem. Biol.* 5 (2009) 108–117.  
23 569 <https://doi.org/10.1038/nchembio.140>.
- 24 570 [36] W. Su, O. Yeku, S. Olepu, A. Genna, J.-S. Park, H. Ren, G. Du, M.H. Gelb, A.J. Morris,  
25 571 M.A. Frohman, 5-Fluoro-2-indolyl des-chlorohalopemide (FIPI), a phospholipase D  
26 572 pharmacological inhibitor that alters cell spreading and inhibits chemotaxis, *Mol. Pharmacol.*  
27 573 75 (2009) 437–446. <https://doi.org/10.1124/mol.108.053298>.
- 28 574 [37] R.R. Lavieri, S.A. Scott, P.E. Selvy, K. Kim, S. Jadhav, R.D. Morrison, J.S. Daniels, H.A.  
30 575 Brown, C.W. Lindsley, Design, synthesis, and biological evaluation of halogenated N-(2-(4-  
31 576 oxo-1-phenyl-1,3,8-triazaspiro[4.5]decan-8-yl)ethyl)benzamides: discovery of an isoform-  
32 577 selective small molecule phospholipase D2 inhibitor, *J. Med. Chem.* 53 (2010) 6706–6719.  
34 578 <https://doi.org/10.1021/jm100814g>.
- 35 579 [38] R. Ganesan, M. Mahankali, G. Alter, J. Gomez-Cambronero, Two sites of action for PLD2  
36 580 inhibitors: The enzyme catalytic center and an allosteric, phosphoinositide binding pocket,  
37 581 *Biochim. Biophys. Acta.* 1851 (2015) 261–272.  
38 582 <https://doi.org/10.1016/j.bbali.2014.12.007>.
- 39 583 [39] S. Majd, E.C. Yusko, A.D. MacBriar, J. Yang, M. Mayer, Gramicidin pores report the activity  
40 584 of membrane-active enzymes, *J. Am. Chem. Soc.* 131 (2009) 16119–16126.  
42 585 <https://doi.org/10.1021/ja904072s>.
- 43 586 [40] J. Tou, C. Urbizo, Resveratrol inhibits the formation of phosphatidic acid and diglyceride in  
44 587 chemotactic peptide- or phorbol ester-stimulated human neutrophils, *Cell. Signal.* 13 (2001)  
45 588 191–197. [https://doi.org/10.1016/s0898-6568\(01\)00137-1](https://doi.org/10.1016/s0898-6568(01)00137-1).
- 46 589 [41] L. Li, N. Fleming, Aluminum fluoride inhibition of cabbage phospholipase D by a phosphate-  
48 590 mimicking mechanism, *FEBS Lett.* 461 (1999) 1–5. [https://doi.org/10.1016/s0014-](https://doi.org/10.1016/s0014-5793(99)01414-3)  
49 591 [5793\(99\)01414-3](https://doi.org/10.1016/s0014-5793(99)01414-3).
- 50 592 [42] M.M. Bradford, A rapid and sensitive method for the quantitation of microgram quantities of  
51 593 protein utilizing the principle of protein-dye binding, *Anal Biochem.* 72 (1976) 248–54.
- 52 594 [43] D.I. Svergun, Restoring low resolution structure of biological macromolecules from solution  
53 595 scattering using simulated annealing, *Biophys. J.* 76 (1999) 2879–2886.  
55 596 [https://doi.org/10.1016/S0006-3495\(99\)77443-6](https://doi.org/10.1016/S0006-3495(99)77443-6).
- 56 597 [44] D.I. Svergun, M.V. Petoukhov, M.H. Koch, Determination of domain structure of proteins  
57 598 from X-ray solution scattering, *Biophys. J.* 80 (2001) 2946–2953.  
59 599 [https://doi.org/10.1016/S0006-3495\(01\)76260-1](https://doi.org/10.1016/S0006-3495(01)76260-1).
- 60  
61  
62  
63  
64  
65

- 1  
2  
3  
4 600 [45] V.V. Volkov, Svergun, D.I., Uniqueness of ab initio shape determination in small-angle  
5 601 scattering, *J. Appl. Crystallogr.* (2003) 860–864.  
6 602 [46] D. Schneidman-Duhovny, M. Hammel, J.A. Tainer, A. Sali, FoXS, FoXSDock and  
7 603 MultiFoXS: Single-state and multi-state structural modeling of proteins and their complexes  
8 604 based on SAXS profiles, *Nucleic Acids Res.* 44 (2016) W424–429.  
9 605 <https://doi.org/10.1093/nar/gkw389>.  
10 606 [47] V. Piiadov, E. Ares de Araújo, M. Oliveira Neto, A.F. Craievich, I. Polikarpov, SAXSMoW  
11 607 2.0: Online calculator of the molecular weight of proteins in dilute solution from experimental  
12 608 SAXS data measured on a relative scale, *Protein Sci. Publ. Protein Soc.* 28 (2019) 454–463.  
13 609 <https://doi.org/10.1002/pro.3528>.  
14 610 [48] Y. Arhab, R. Rahier, A. Noiriél, M.V. Cherrier, A. Abousalham, Expression and Purification  
15 611 of Recombinant *Vigna unguiculata* Phospholipase D in *Pichia pastoris* for Structural Studies,  
16 612 in: G. Sandoval (Ed.), *Lipases Phospholipases*, Springer New York, New York, NY, 2018:  
17 613 pp. 191–201. [https://doi.org/10.1007/978-1-4939-8672-9\\_10](https://doi.org/10.1007/978-1-4939-8672-9_10).  
18 614 [49] R. Rahier, A. Noiriél, A. Abousalham, Development of a Direct and Continuous  
19 615 Phospholipase D Assay Based on the Chelation-Enhanced Fluorescence Property of 8-  
20 616 Hydroxyquinoline, *Anal. Chem.* 88 (2016) 666–74.  
21 617 <https://doi.org/10.1021/acs.analchem.5b02332>.  
22 618 [50] J. Zhang, T. Chung, K. Oldenburg, A Simple Statistical Parameter for Use in Evaluation and  
23 619 Validation of High Throughput Screening Assays, *J. Biomol. Screen.* 4 (1999) 67–73.  
24 620 <https://doi.org/10.1177/108705719900400206>.  
25 621 [51] H. Sato, T. Watanabe, Y. Sagane, Y. Nakazawa, K. Takano, Purification and Characterization  
26 622 of Phospholipase D from Cabbage Leaves., *Food Sci. Technol. Res.* 6 (2000) 29–33.  
27 623 <https://doi.org/10.3136/fstr.6.29>.  
28 624 [52] Y. Nakazawa, H. Sato, M. Uchino, K. Takano, Purification, characterization and cloning of  
29 625 phospholipase D from peanut seeds, *Protein J.* 25 (2006) 212–223.  
30 626 <https://doi.org/10.1007/s10930-006-9004-9>.  
31 627 [53] S. Stumpe, S. König, R. Ulbrich-Hofmann, Insights into the structure of plant  $\alpha$ -type  
32 628 phospholipase D: Structure of plant phospholipase D, *FEBS J.* 274 (2007) 2630–2640.  
33 629 <https://doi.org/10.1111/j.1742-4658.2007.05798.x>.  
34 630 [54] Arhab, Y, Rahier, R, Noiriél, A, Cherrier, MV, Abousalham, A, Expression and Purification  
35 631 of Recombinant *Vigna unguiculata* Phospholipase D in *Pichia pastoris* for Structural Studies,  
36 632 *Methods Mol. Biol. Press.* (2018) In press.  
37 633 [55] K. Pappan, X. Wang, Molecular and biochemical properties and physiological roles of plant  
38 634 phospholipase D, *Biochim. Biophys. Acta.* 1439 (1999) 151–166.  
39 635  
40  
41  
42  
43  
44  
45  
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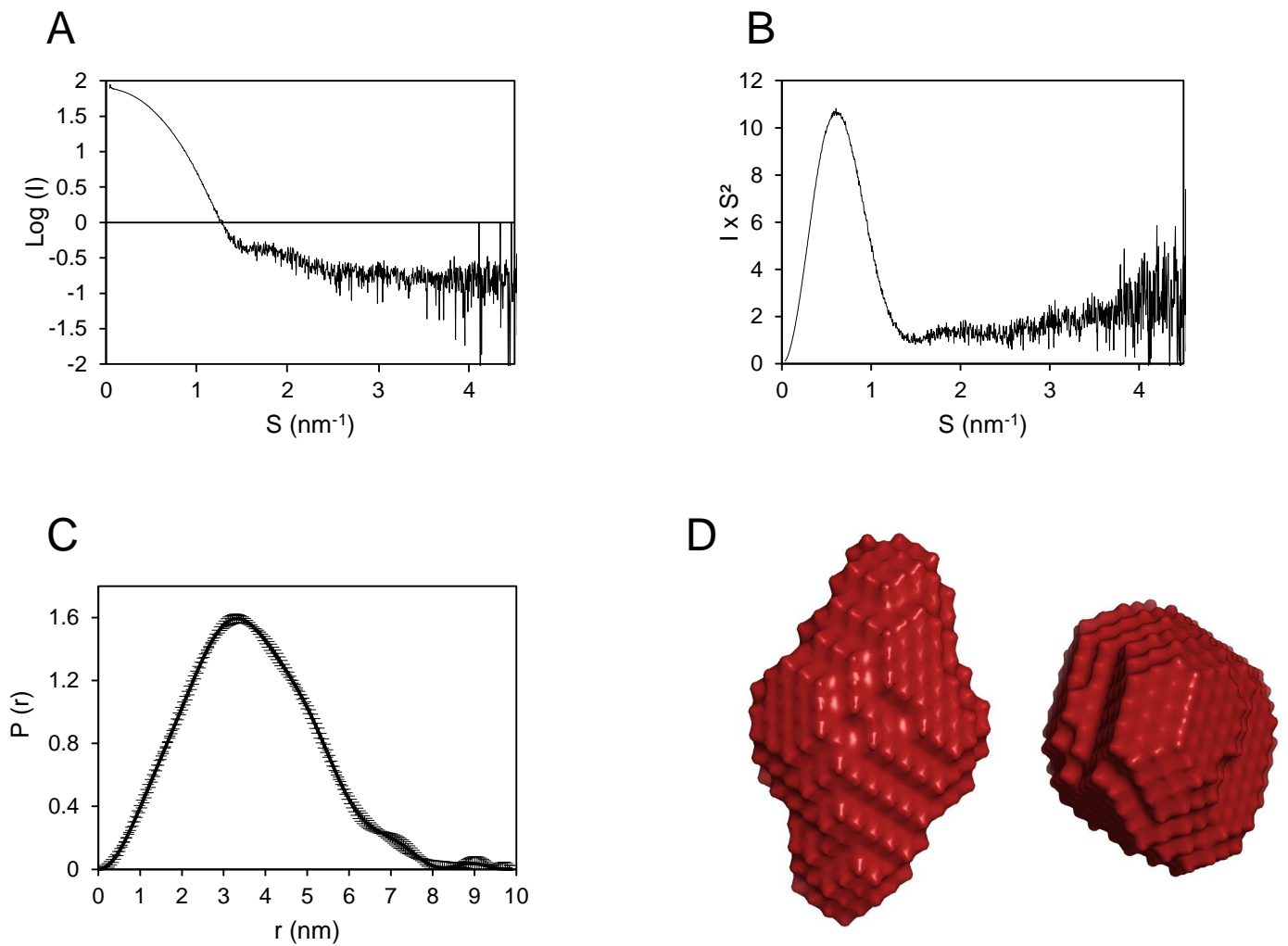


Fig. 1: SAXS modelization of VuPLD.

A) Diffusion curve. B) Kratky curve. C) Distance distribution function  $P(r)$  calculated from X-ray scattering data. D) *Ab initio* low-resolution structure model of VuPLD calculated from the SAXS pattern and the program DAMMIN. Model is shown in two different views obtained by  $90^\circ$  rotation around the y-axis.

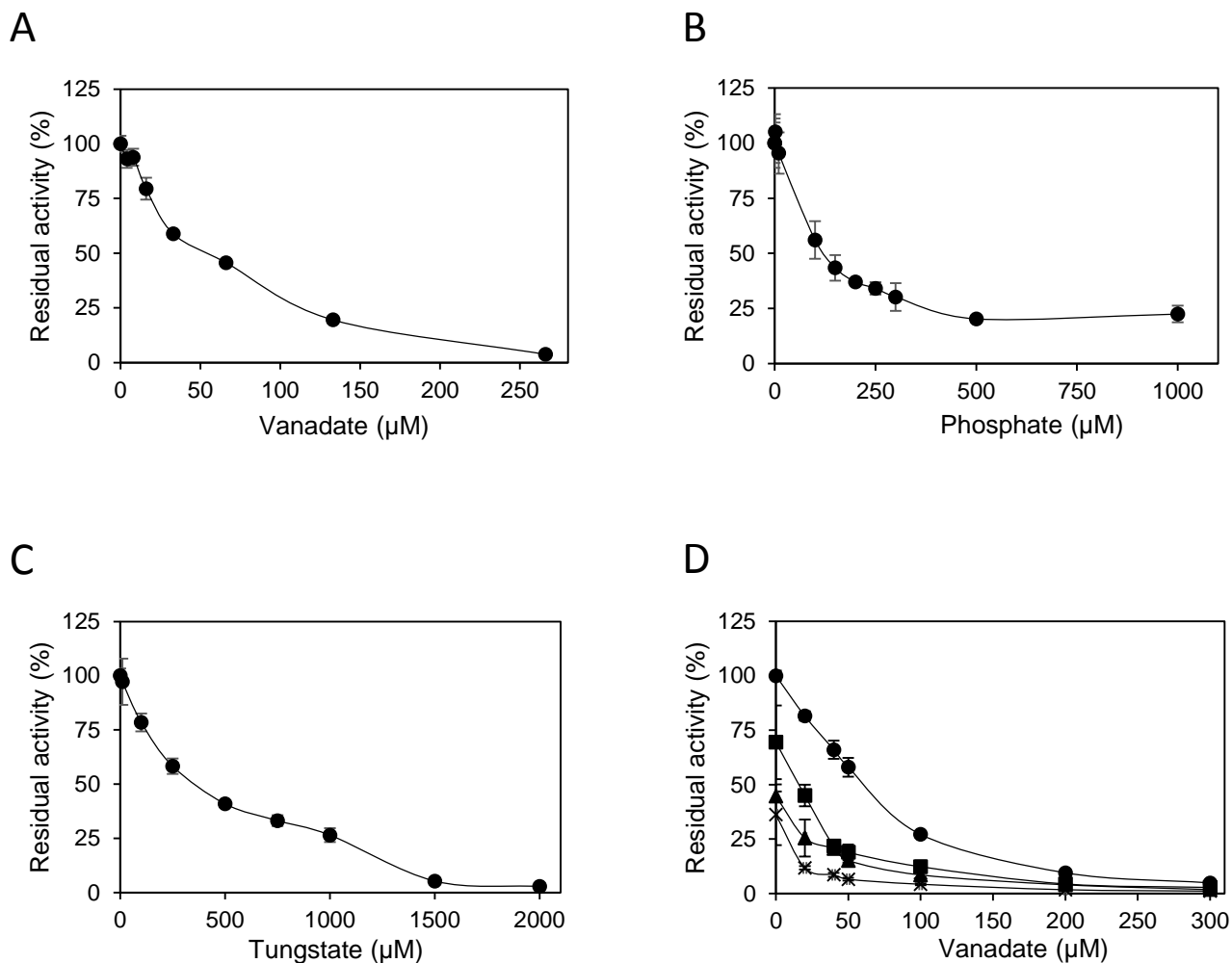


Fig. 2: Inhibitory effect of different salts on purified VuPLD.

The PLD activity was measured with the fluorimetric assay in triplicates and with increasing concentrations of sodium vanadate (A), sodium tungstate (B) or sodium phosphate (C). D) PLD relative activity incubated with 0  $\mu\text{M}$  (circle ●), 25  $\mu\text{M}$  (square ■), 50  $\mu\text{M}$  (triangle ▲) or 100  $\mu\text{M}$  (cross \*) of phosphate at different concentrations of vanadate. Relative PLD activities were calculated relatively to the enzyme incubated in the absence of effector set at 100 %. Values are the mean  $\pm$  SD obtained from three independent assays.

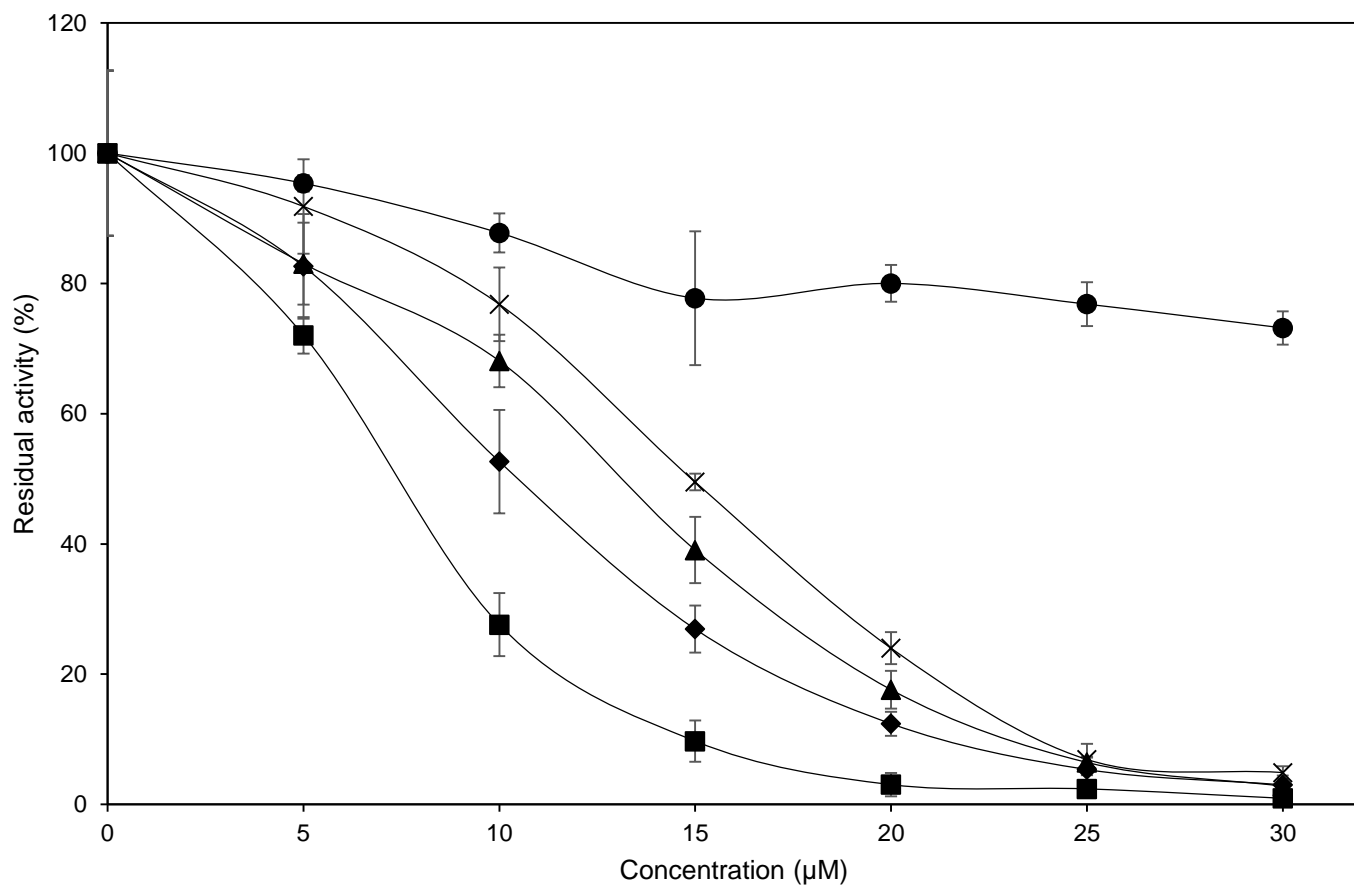


Fig. 3: Inhibitory effect of different vanadate derivatives on purified VuPLD.

The activity was measured with the chromogenic assay in triplicates with increasing concentrations of vanadate (circle ●), vanadate oxytriethoxide (square ■), vanadate oxytripropoxide (triangle ▲), vanadate oxytriisopropoxide (cross ×) or vanadate tris(triphenylsiloxy) (diamond ◆). Values are the mean  $\pm$  SD obtained from three independent chromogenic assays.

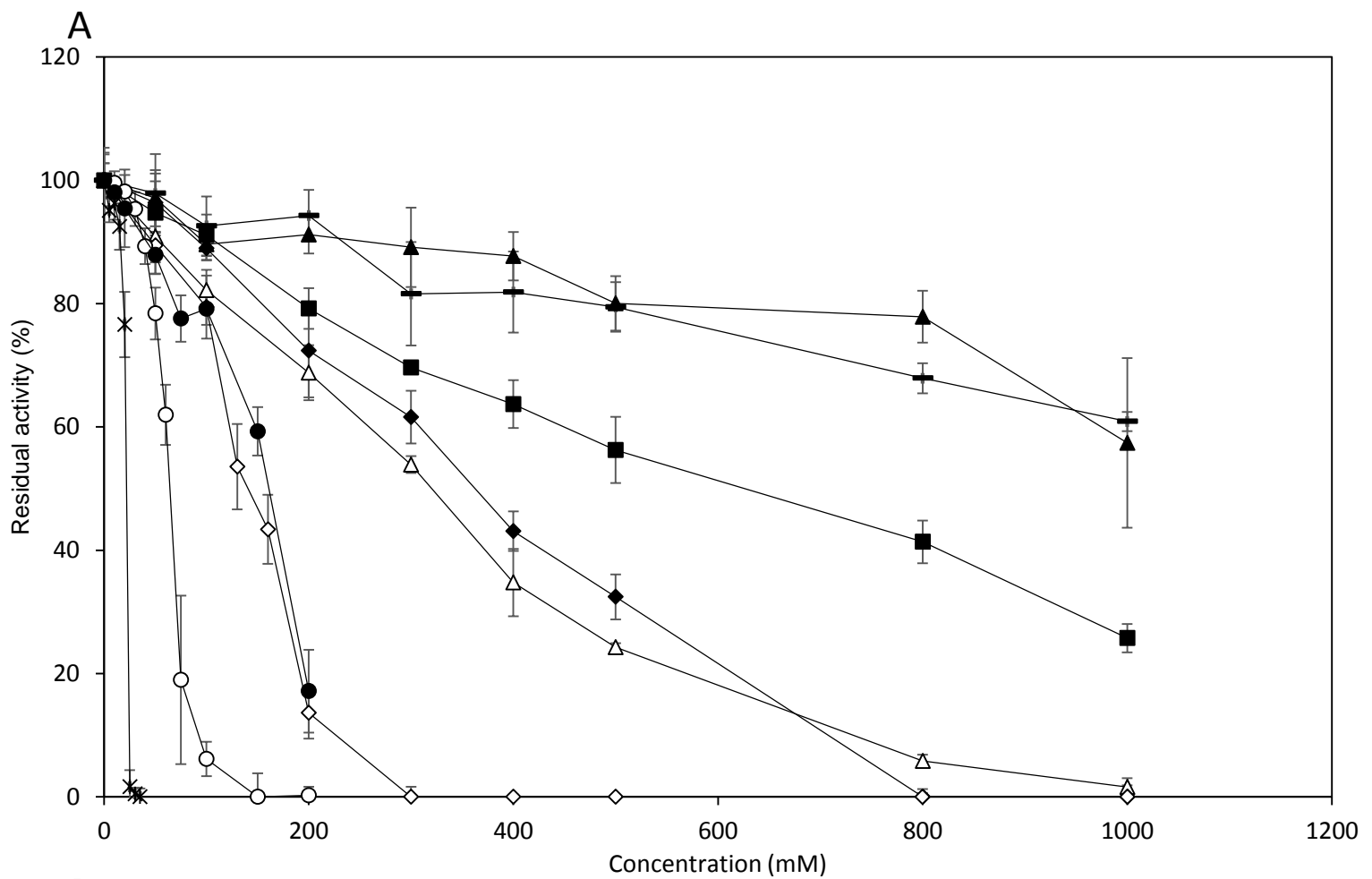


Fig. 4: Effect of different alcohols on purified VuPLD.

A) The activity was measured in triplicates with increasing concentrations of methanol (black dash—), ethanol (square■), propan-1-ol (white triangle Δ), propan-2-ol (black triangle ▲), butan-1-ol (white diamond ◇), butan-2-ol (black diamond ◆), pentan-1-ol (white circle ○), pentan-2-ol (black circle ●) and hexanol (cross \*). Relative activities were calculated relatively to the enzyme incubated in the absence of effector (100%). Values are the mean  $\pm$  SD obtained from three independent fluorimetric assays. B) TLC analysis of the VuPLD catalyzed transphosphatidylation reaction products obtained from (A). Lipids were extracted, developed, and revealed as described in the experimental section. POPA, phosphatidylbutanol (P-But) and phosphatidylethanol (P-Eth) were run as standard reaction products,



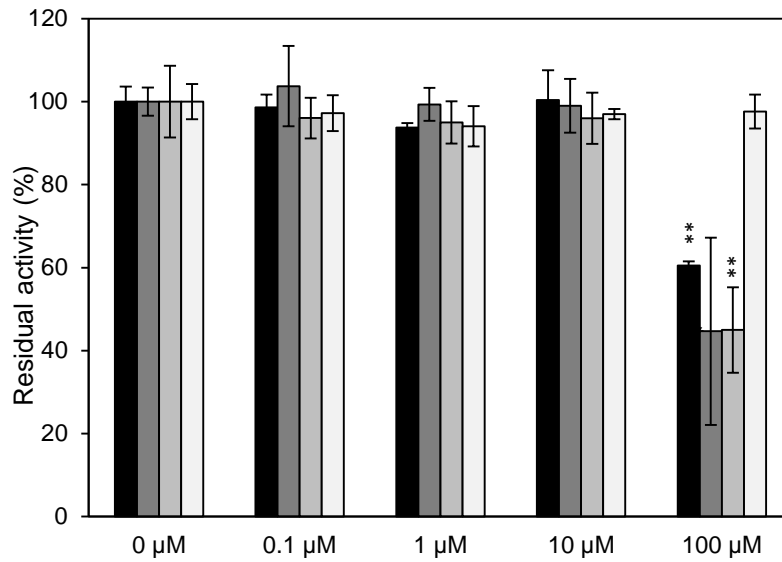


Fig. 5: Inhibitory effect of known inhibitors of human PLD on purified plant VuPLD.

The activity was measured in triplicates with increasing concentrations of halopemide (black), VU0359595 (dark grey), VU0285655 (light grey), FIPI (white). Relative activities are calculated relatively to the enzyme incubated with 0 μM of effector (100%). Values are the mean ± SD obtained from three independent fluorimetric assays. \*\* P<0.01, \*\*\* P<0.001 (vs untreated).

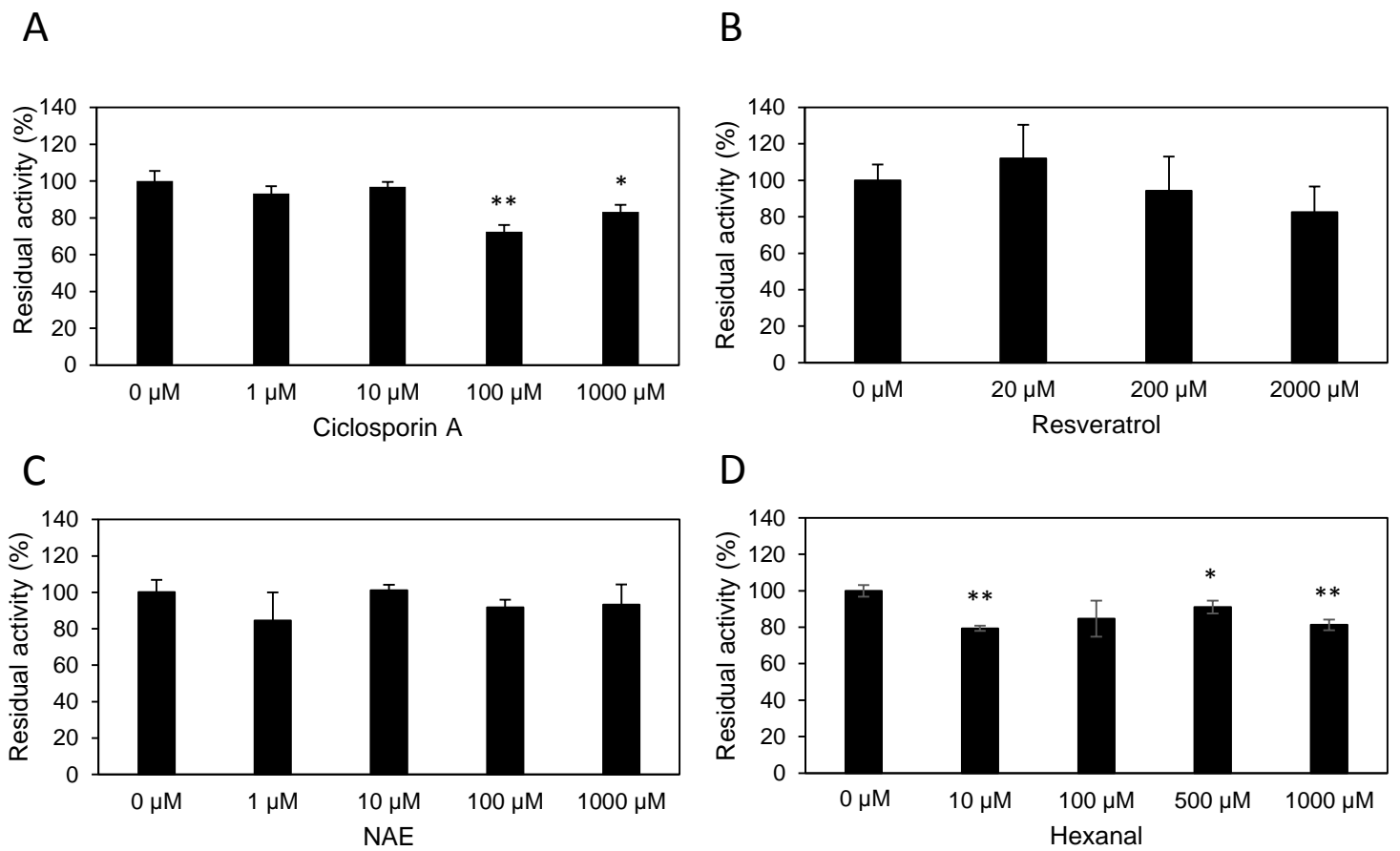


Fig. 6: Inhibitory effect of alleged inhibitors of plant PLD.

PLD described inhibitors, ciclosporin A (A), resveratrol (B), NAE C12:0 (C) and hexanal (D) were tested at different concentrations from 1  $\mu\text{M}$  to 1 mM. Relative activities are then calculated relatively to the enzyme incubated in the absence of effector (100%). Values are the mean  $\pm$  SD obtained from three independent fluorimetric assays. \* $P < 0.05$ , \*\* $P < 0.01$  (vs untreated).

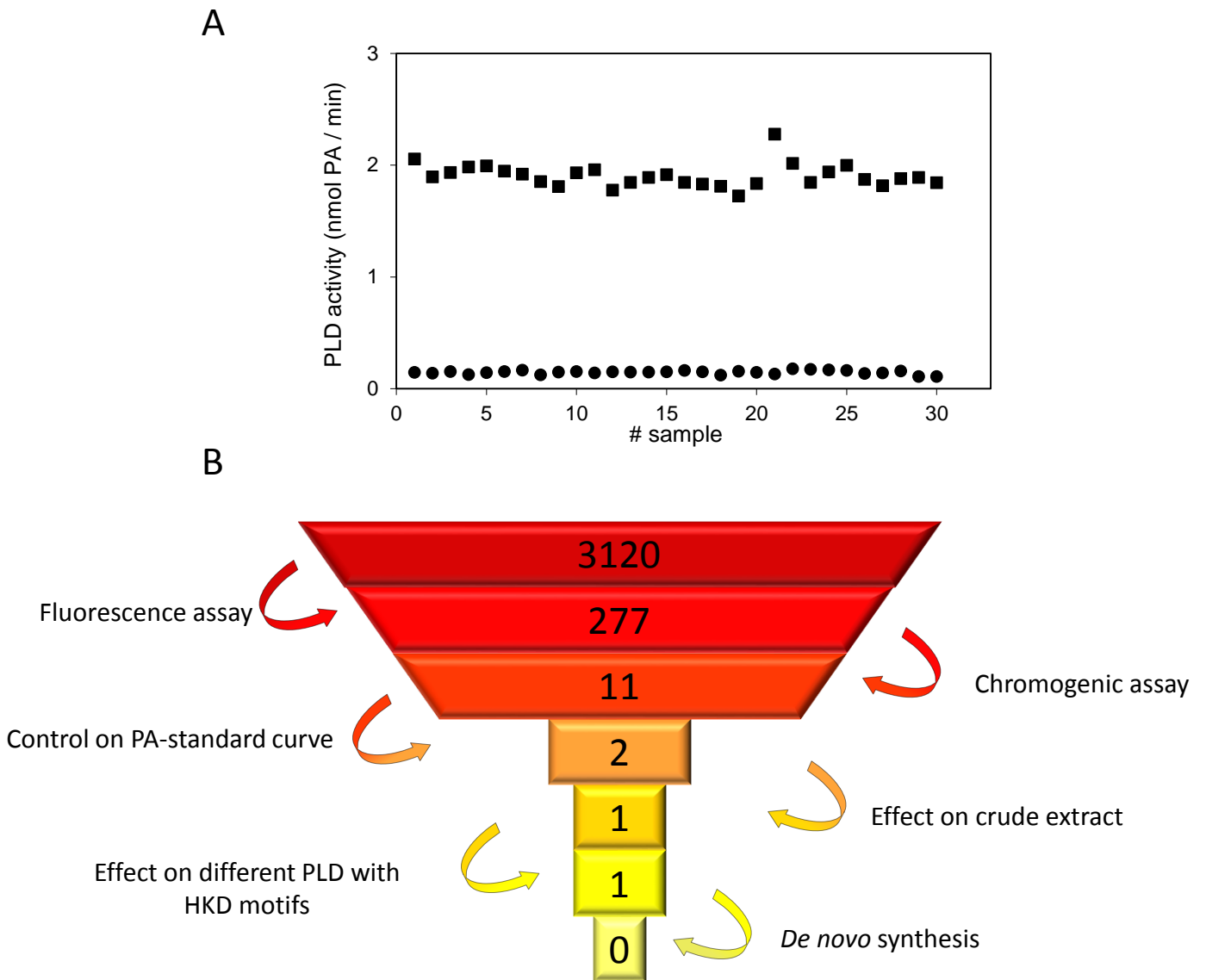


Fig. 7: HTS screening of a new PLD inhibitor.

PLD activity was measured in the presence or absence of 100  $\mu\text{M}$  of vanadate in order to establish the variation of the assay and the calculation of the Z' score. Multiple repeats ( $n=30$ ) were realized with (circles) or without effector (squares). B) Seven-step workflow for the identification of novel PLD inhibitors.

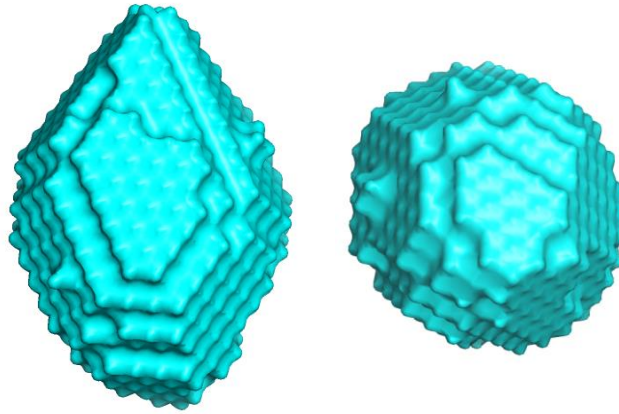


Fig. S1: SAXS modelization of VuPLD.  
*Ab initio* low-resolution structure model of VuPLD calculated from the SAXS pattern and the program GASBOR. Model is shown in two different views obtained by 90° rotation around the y-axis

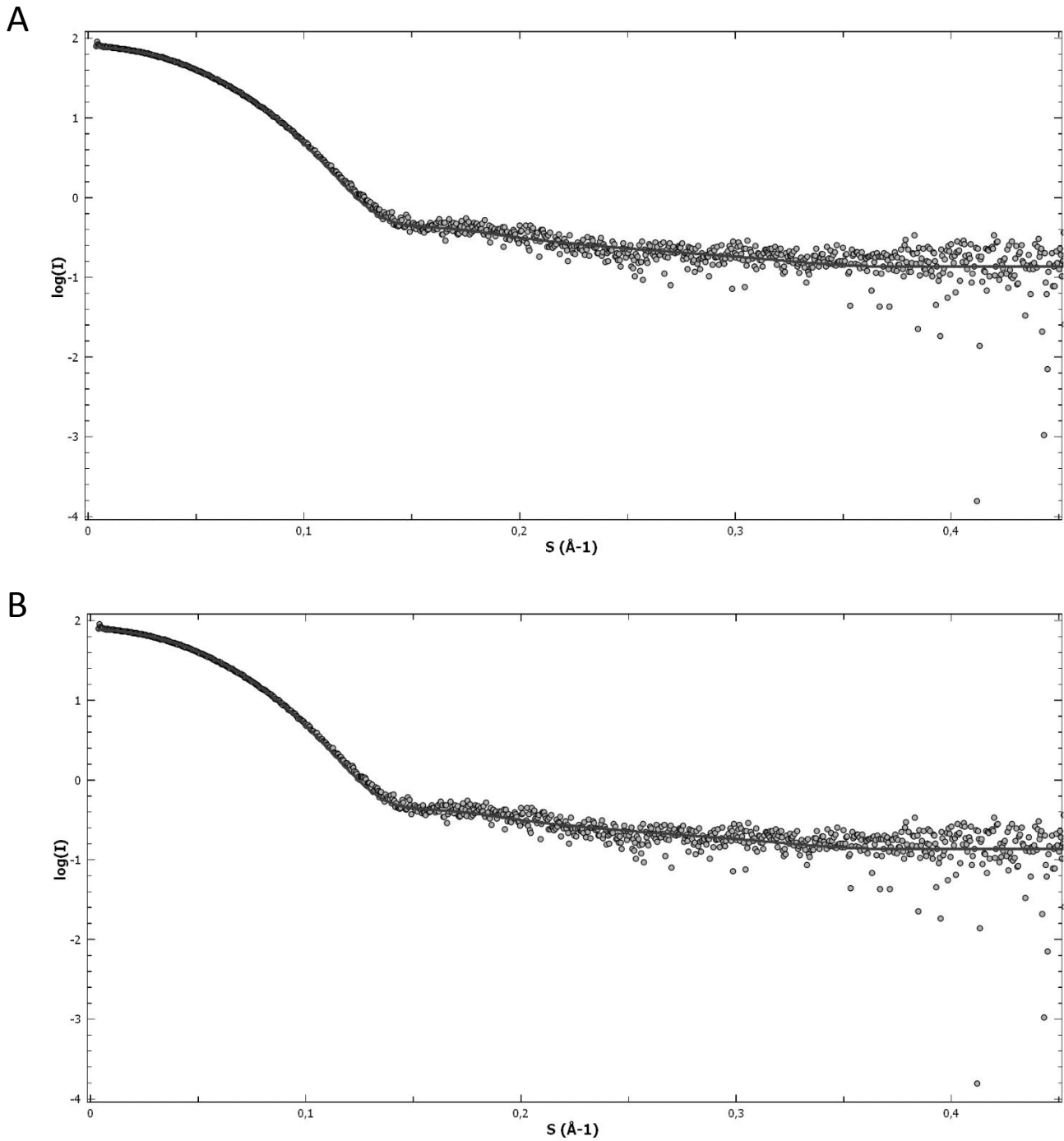


Fig. S2: Analysis of SAXS data of VuPLD.

A) Comparison of the experimental scattering data (gray circles) with the calculated scattering profile from PLD structure.

B) Experimental SAXS curve of VuPLD (grey dots) compared to the theoretical SAXS curve of PLD $\alpha$ 1 (black line) from Arabidopsis (PDB code 6KZ9).

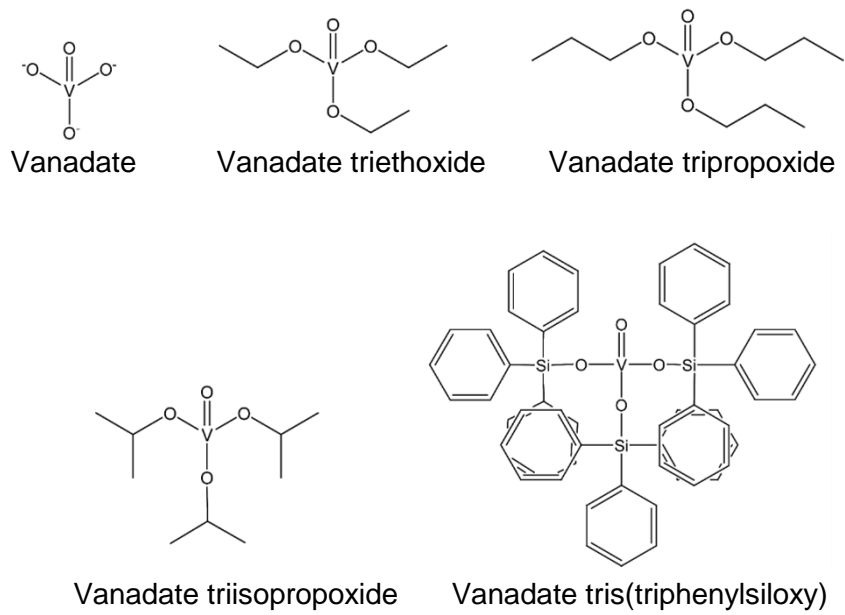


Fig. S3: Chemical structures of vanadate and its derivatives

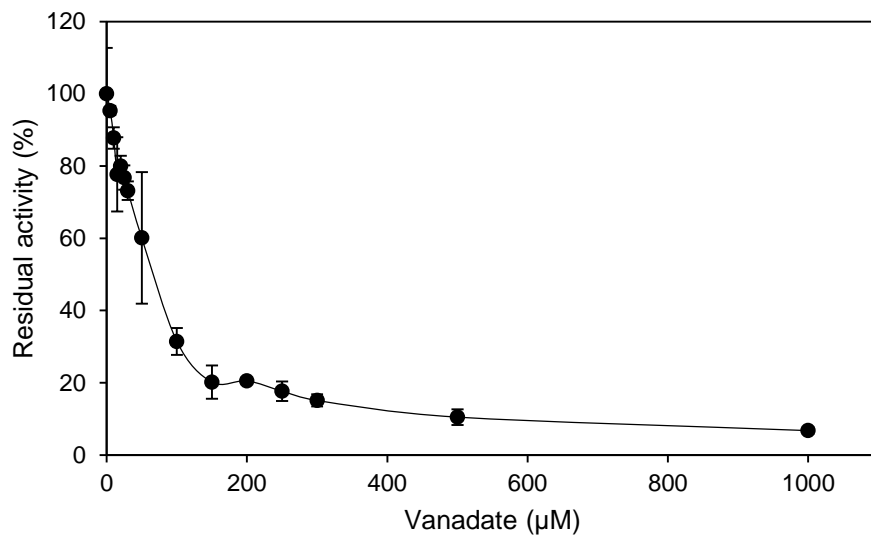


Fig. S4: Inhibition of purified plant PLD by vanadate. The activity was measured in triplicates with increasing concentrations of vanadate. Values are the mean  $\pm$  SD obtained from three independent chromogenic assays.

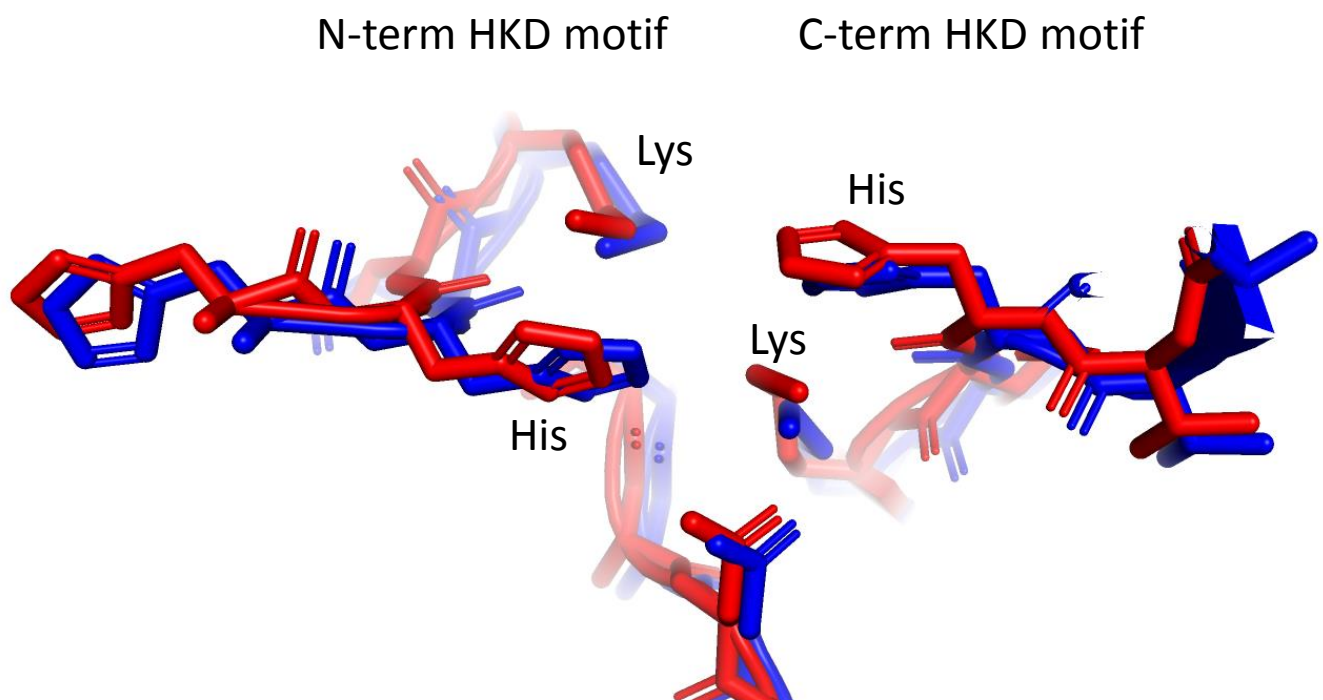


Fig. S5: Comparison of plant and mammalian PLD catalytic pockets. Both HKD motifs are displayed in blue (human PLD, 6OHR) and red (plant PLD, 6KZ9) according to [9]. Histidyl and lysyl catalytic residues are represented (sticks) and annotated. Structure superposition was done using Pymol 2.3.2..



## **Author Statement**

YA and MVC performed SAXS and modeling experiments. AC and FP contributed to compound design and synthesis. YA, KB, HA, MA, RR performed biochemical assays. LB, SM, AA and AN devised the project and the main conceptual ideas. AN was in charge of funding acquisition. All authors contributed to data interpretation. The manuscript was written from contributions by all the authors. All the authors have given approval to the final version of the manuscript.